

Quantification of Cocaine Hydrochloride in Seized Drug Samples by Infrared Spectroscopy and PLSR

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A determinação do teor de cocaína em amostras de drogas consiste em uma importante tarefa em órgãos como a Polícia Federal Brasileira (BFP). Nesse sentido, este trabalho propõe apresentar um método baseado em espectros de infravermelho obtidos por refletância total atenuada (ATR) e regressão por mínimos quadrados parciais (PLSR) para quantificar cloridrato de cocaína em amostras de drogas. O método foi desenvolvido e validado com 275 amostras reais de drogas apreendidas pela BFP em todo o Brasil. A determinação foi realizada no intervalo de 35 a 99% (m/m) de cocaína nas amostras. Os resultados indicaram que o método é capaz de analisar diretamente amostras de drogas contendo cocaína na forma de cloridrato sem necessidade de qualquer preparo de amostra com erros médios de aproximadamente 3,00%, precisão de 1,50% (m/m) e concentração mínima detectável de 13% (m/m).

The determination of cocaine in drug samples is an important task for law enforcement agencies such as the Brazilian Federal Police (BFP). In this sense, this paper proposes a method based on infrared spectra obtained by attenuated total reflectance (ATR) and partial least squares regression (PLSR) to quantify cocaine hydrochloride in drug samples. The method was developed and validated with 275 actual samples of drugs seized by the BFP. The determination was performed between 35 to 99% (m/m) of cocaine in the drug samples. Results indicate that the method is able to directly analyze drug samples containing cocaine in its hydrochloride form without any sample preparation with average prediction errors of 3.00% (m/m), 1.50% (m/m) precision and 13% (m/m) of minimum detectable concentration.

Keywords: cocaine hydrochloride, PLSR, FTIR

Introduction

According to the United Nations Office on Drugs and Crime (UNODC), cocaine is the second most problematic drug worldwide in terms of negative health consequences, and probably the most problematic one in terms of drug trafficking-related violence.¹ Furthermore, a World Drug Report published in 2013 asserts that while the use of cocaine in several countries in South America decreased or remained stable, in Brazil it has increased substantially, which justifies the intensification of studies that can assist law enforcement agencies in the control of illicit drug trafficking.²

In order to increase drug volumes and illegal trafficking profits, various substances are added to cocaine. Among

them there are diluting agents such as sugars and starches, as well as adulterants, which are pharmacologically active compounds capable of increasing the drug's adverse and side effects.¹ The determination of cocaine concentration and of its diluents and adulterants has a significant role in forensic sciences, since it provides important information on how the drug has been cut, as well as on how illegal distribution networks operate in a certain area.

Since the 1970s, the analytical methods used for the determination of cocaine in drug samples have evolved substantially.³ Currently, gas chromatography is the main technique applied for this analysis, since it provides accurate and precise results.^{4,5} Furthermore, gas chromatography is the recommended technique in the UNODC's manuals.⁶⁻⁸ However, despite the excellent results, gas chromatography usually requires a relatively complex sample preparation procedure and results in high

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cost and time for analysis. In addition, sample diversity demands the method to be frequently adjusted or updated. These factors make the analysis of a large number of drug samples by gas chromatography time consuming and difficult to implement.

Multivariate analysis methods based on Fourier transform infrared (FTIR) spectroscopy have shown great potential for both qualitative and quantitative analyses. The combination of vibrational spectroscopy and chemometrics has been considered an efficient alternative method to directly extract information from many different data.⁹ One of the first methods applying FTIR for cocaine determination was proposed by Ravreby.¹⁰ In that case, cocaine hydrochloride and heroin concentrations were determined by univariate regression choosing a carbonyl absorption peak in the infrared (IR) spectra obtained with KBr pellets. The author also studied the effect of various additives and diluents such as starch, sugars, mannitol, caffeine, and procaine. However, considering the high complexity of seized drugs and the variation of the IR spectra obtained with KBr pellets, this method presents limitations for routine application in forensic analysis.¹⁰ Ryder *et al.* showed that multivariate analysis methods combined with Raman spectroscopy can be used as a rapid analytical method for the analysis of narcotics in two component mixtures. In this study, partial least squares regression (PLSR) showed that Raman data allow the estimation of cocaine concentration in solid mixtures with glucose, which should be sufficient for screening of samples.¹¹ Rodrigues *et al.* performed an exploratory study that characterized the chemical composition of 91 cocaine samples seized in the state of Minas Gerais between 2008 and 2010, based on attenuated total reflectance (ATR) FTIR spectra and chemometric analysis. In their study, principal component analysis (PCA) and partial least squares for discriminant analysis (PLS-DA) were developed to classify the samples according to their dilution (below and above 15% m/m) and chemical form (cocaine hydrochloride or base). Discrimination according to dilution and chemical form resulted respectively in 83% and 97% correct results.⁹ In a preliminary study, Maharaj compared the quantification of cocaine using gas chromatography and flame ionization detection (GC-FID) with FTIR.¹² Although only few samples were used for analysis, and the lack of results for figures of merit of the method, the author affirmed that the results obtained by GC-FID and ATR-FTIR are equivalent and consequently, ATR-FTIR was considered adequate for the quantitative analysis of cocaine.¹² Recently, Pérez-Alfonso *et al.* proposed a method for determination of cocaine in illicit samples by diffuse reflectance measurements in the near infrared

spectroscopy (NIR) region. The results obtained by the authors showed that the cocaine content can be determined in a wide concentration range. However, the validation was performed with a relatively low number of samples if one takes into account the high heterogeneity of illicit samples occurring in real forensic cases.¹³

The results described in the literature suggest that infrared spectroscopy and multivariate analysis can be a viable analytical method for cocaine analysis in drug samples. However, there is still a lack of validated methods with a significant number of seized cocaine samples in order to prove the real potential of this technique in routine forensic analysis. Therefore, the aim of the present work is to describe the development and validation of an analytical method to quantify cocaine hydrochloride in seized drug samples using ATR-FTIR and multivariate calibration. In order to develop and validate the proposed method, the quantitative determination of cocaine was performed in a significant number of drug samples seized nationwide using two independent analytical techniques, namely GC-FID and ATR-FTIR.

Experimental

Samples and sample preparation

The dataset was composed of 275 samples of cocaine hydrochloride originating from approximately 73 seizures made by the Brazilian Federal Police (BFP) in different parts of Brazil, between 2009 and 2013. All samples were sent to the Forensic Chemistry Laboratory of the National Institute of Criminalistics in Brasília. Before instrumental analysis, all samples were carefully homogenized by maceration.

Infrared spectroscopy measurements

The infrared spectra were obtained on a Nicolet iS10 FTIR spectrometer equipped with a triple reflection attenuated total reflectance SMART iTR accessory using a diamond crystal. The measurements were obtained in reflectance mode (R) with the accessory filled with a small amount of cocaine. The spectra were collected between 4000 and 400 cm^{-1} over 16 scans with a resolution of 4.0 cm^{-1} .

Gas chromatography with flame ionization detection (GC-FID) analysis

GC-FID was used as the reference method. The cocaine content in the drug samples was determined by weighing

an amount of 12.25 mg \pm 0.25 mg of each homogenized sample and mixing thoroughly with 10.0 mL of an internal standard solution (diethylamine, 0.002 mL L⁻¹ and dipentyl phthalate, 512 mg L⁻¹ prepared in chloroform). Then, 1 mL of this solution was transferred to glass vials, sealed, and subjected to chromatographic separation.

GC-FID analysis was performed on a gas chromatograph model 6890N (Agilent Technologies) equipped with a flame ionization detector and an autosampler 7683B Series (Agilent Technologies). The chromatographic conditions were as follows: injection volume of 0.2 mL; split ratio of 50:1; chromatographic column DB1-MS methyl siloxane (25 m \times 200 μ m [i.d.] \times 0.33 μ m film thickness); injector temperature of 280 °C, and detector temperature of 320 °C. Helium was used as the carrier gas at a flow of 1.0 mL min⁻¹. The oven temperature program was as follows: 150 °C for 2 min, heat 40 °C min⁻¹ to 350 °C, and hold at 350 °C for 4.5 min, resulting in a 12 min chromatographic run.

Multivariate model development

The multivariate calibration method was developed based on partial least squares regression (PLSR). In PLSR, the original spectral variables are decomposed into latent variables in order to establish the best correlation between the instrumental measurements (spectral data matrix X) and the values of the interest property (vector y containing the reference values of cocaine concentration).¹⁴⁻¹⁶

All sample spectra were imported into MATLAB® (version 7.12, R2011a) and the preprocessing and PLSR models were implemented using the PLS Toolbox® (version 6.5) from Eigenvector Technologies. Two independent regression models were developed. The first one was established using the data on a reflectance scale, while the second one utilized converted absorbance (Abs) data by means of the relation Abs = log₁₀(1/R).

To perform the calibration and validation of the models, the dataset was split respectively into 184 calibration and 91 validation samples selected by the Kennard-Stone algorithm.¹⁷

In order to obtain the best prediction results, several pre-processing techniques were evaluated, specifically standard normal variate (SNV), orthogonal signal correction (OSC), first derivative, mean center, and their combinations. The selection of the best preprocessing method was made based on the root mean square error of calibration (RMSEC) and the mean error of cross validation (RMSECV) obtained by 92 continuous blocks.^{14,15}

After choosing the pre-processing method, the models were optimized by the elimination of outliers. Methods for outlier identification have been described in detail in

several publications.^{15,16} In this work, outlier identification was performed as described in ASTM E1655-05¹⁸ and in the references published by Valderrama *et al.*, based on data with extreme leverage, unmodeled residuals in spectral data and unmodeled residuals in the dependent variable, taking into account 99% confidence intervals.¹⁹ Initially, a first calibration model was built and the outliers were removed from the calibration samples; then the model was recalculated and the outlier identification and exclusion process was repeated. After two outlier exclusions, the third model was considered to be optimized. The validation set was evaluated with the optimized calibration model and the outliers were excluded by applying the same criteria used for the calibration model.

Analytical figures of merit

Trueness is the parameter that informs the degree of agreement between the reference and the estimated values by the proposed method.²⁰ In average terms, it can be expressed as the root mean square error of prediction (RMSEP), which is an approximation of the average prediction error for the validation samples obtained from equation 1.²¹

$$\text{RMSEP} = \sqrt{\frac{\sum_{i=1}^{I_V} (y_i - \hat{y}_i)^2}{I_V}} \quad (1)$$

where I_V is the number of validation samples, while y_i and \hat{y}_i are respectively the reference value and estimated value for the cocaine concentration for sample i . Another parameter used to measure the degree of agreement between the reference value and estimated value is the relative error of prediction (REP), which is determined by equation 2.²¹

$$\text{REP} = 100 \sqrt{\frac{\sum_{i=1}^{I_V} (y_i - \hat{y}_i)^2}{I_V y_i^2}} \quad (2)$$

The sensitivity (SEN) of the method determines the fraction of the analytical signal due to the increase in the concentration of a particular analyte in the unit concentration. The SEN was determined based on the regression coefficients of the PLSR model, according equation 3.²¹⁻²³

$$\text{SEN} = \frac{1}{\|b\|} \quad (3)$$

where b is the vector of regression coefficients with A latent variables.

The precision of the method measures the dispersion of estimated results for the interest property obtained from independent experiments which are repeated for a same sample under the defined conditions.^{24,25} Precision was determined according to equation 4. For this purpose, four different samples with concentrations regularly distributed along the linear range of the method, with 10 replicates each performed in the same day, were analyzed.

$$\text{Precision} = \sqrt{\frac{\sum_{i=1}^I \sum_{j=1}^m (\hat{y}_{i,j} - \hat{\bar{y}}_i)^2}{I(m-1)}} \quad (4)$$

where I is the number of samples, m is the number of replicates, $\hat{y}_{i,j}$ is the estimate concentration for sample i and replicate j , and $\hat{\bar{y}}_i$ is the average concentration of the replicates for sample i .

The minimum detectable concentration (MDC) is defined as the lowest concentration that can be reliably measured. The MDC can be determined by applying ISO 11843-2 recommendations.^{26,27} Ortiz *et al.* suggests that the proposed ISO 11843-2 calculation can be directly extended to multivariate cases.²⁸ Thus, the MDC values were calculated as suggested by Ortiz *et al.* using equation 5.²⁸

$$\text{MDC} = \delta_{\alpha,\beta,\nu} \frac{s}{b} \sqrt{\frac{1}{m} + \frac{1}{I_C} + \frac{\bar{y}^2}{\sum_{i=1}^{I_C} (y_i - \bar{y})^2}} \quad (5)$$

where s is the standard deviation of the residues for the linear regression between reference values and estimated values by the proposed method, b is the slope of the regression line, I_C is the number of calibration samples, m is the number of replicates, \bar{y} is the median concentration in the calibration samples, $\delta_{\alpha,\beta,\nu}$ is the non-centrality parameter of the t distribution, α and β are the probabilities of occurrence for false negative and false positive errors, respectively, and $\nu(\text{MDC}) = I_C - 2$ degrees of freedom. In this work, the two probabilities α and β were considered to be equal to 0.05 (95% confidence level). It should be noted that the MDC estimated by equation 5 might be considered as an average to all possible MDC for future test samples, since the detectability of the PLSR model depends on the level of other background constituents.²⁹

The confidence intervals can be defined as a range, with a given degree of confidence (i.e., a certain probability) that the real value for the concentration of the analyte of interest is included. This can be determined by applying a residual distribution model (usually the normal distribution) and the estimated standard error of prediction ($s(\hat{y} - y)$), which are determined by equations 6 and 7, respectively.^{23,30}

$$CI(y_i) = \hat{y}_i \pm t_{\nu,1-\alpha/2} \times s(\hat{y} - y) \quad (6)$$

$$CI(y_i) = \hat{y}_i \pm t_{\nu,1-\alpha/2} \times \sqrt{MSEC(1 + h_i + 1/I_C)} \quad (7)$$

where α is the significance level equal to 0.05 (95% confidence level); $t_{\nu,1-\alpha/2}$ is the corresponding critical level for the Student's t distribution with ν pseudo degrees of freedom, determined as proposed by Van der Voet;³¹ $MSEC$ is the mean square error estimated in the calibration samples with ν pseudo degrees of freedom; and h_i is the leverage of the sample, estimated by equation 8.³⁰

$$h_i = t_i^T (T^T T)^{-1} t_i \quad (8)$$

where t_i and T are respectively the scores for sample i and for all the calibration samples, respectively.

Results and Discussion

Figure 1 shows the ATR-FTIR spectra obtained for (a) cocaine hydrochloride standard and (b) all the calibration samples, expressed in reflectance units. According to Rodrigues *et al.*, the spectral region that lies around 2540 cm^{-1} is characteristic of cocaine hydrochloride, attributed to the N–H stretching due to the hydrochloride salt formation.⁹ However, a high number of infrared signals is observed in the infrared spectra of the cocaine standard, most of them also present in the calibration samples. Several infrared bands may be nominated: 729 , 1026 and 1071 cm^{-1} (corresponding to the out-of-plane bending and the mono substituted benzene stretching); 1105 , 1265 and 1230 cm^{-1} (acetate C–O stretching); the bands between 1490 – 1460 cm^{-1} (C–H bending vibrations) and the bands at 1712 and 1728 cm^{-1} (stretching vibration of the two carbonyl groups). Furthermore, it is observed in Figure 1b a significant spectral variation in the data, which can be attributed to differences in cocaine content, the presence of diluting/adulterant agents, and instrumental variations.

In Figure 2, the regression vector of the PLSR model developed with reflectance data is presented. It can be observed that the regression coefficients with the highest absolute values correspond to wavenumbers between 500 and 800 cm^{-1} and between 1400 and 1800 cm^{-1} . The last region can be attributed to the stretching vibration of the two carbonyl groups at 1728 and 1712 cm^{-1} and the C–H bending vibrations at 1490 and 1460 cm^{-1} .

Table 1 shows the results for each PLSR model developed by excluding outliers and the variation of RMSEC and RMSEP values in these models. In both the reflectance and absorbance models, a significant decrease of the RMSEC values is observed after outlier exclusion.

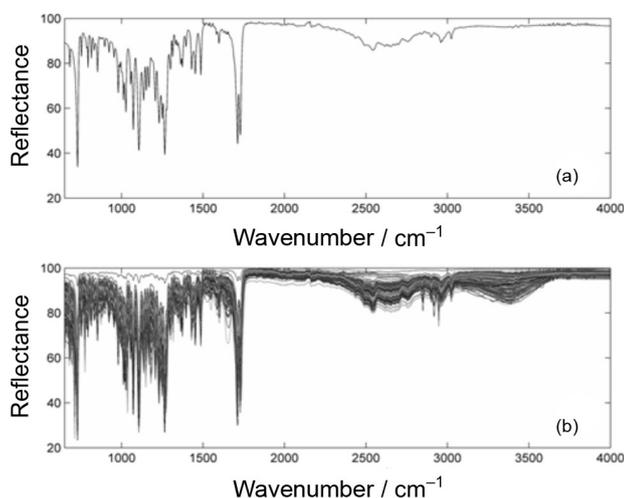


Figure 1. ATR-FTIR spectra of (a) cocaine hydrochloride standard and (b) the 184 drug samples used for model development.

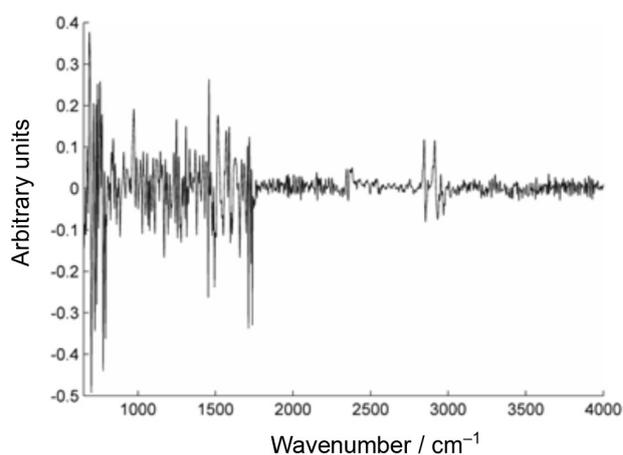


Figure 2. Regression coefficients for the PLSR model developed for reflectance data.

In addition, although some samples have been identified as outliers based on the leverage criterion in the third model (after the second exclusion), they were not excluded from the dataset since the ASTM E1655-05 indicates that the data may be presenting the “snowball effect”. In these cases, the ASTM E1655-05 suggests that the leverage criterion can be relaxed provided that no calibration samples have a leverage greater than 0.5. In these datasets, the high leverage observed in the optimized models for the reflectance and absorbance data was 0.20 and 0.23, respectively.¹⁸

Table 1 also shows that when the model was built based on reflectance and absorbance measurements, 15.8% and 14.1% of the calibration samples and 7.7% and 6.6% of the validation samples were excluded, respectively. However, considering the high heterogeneity of the drug

samples, the number of outliers excluded in the calibration and validation samples was considered to be acceptable in both optimized PLSR models. Samples identified as outliers were analyzed in detail to verify the reasons for their exclusion. More than 60% of the outliers were part of seizures carried out at least three years ago, which may be the cause of changes in some of the chemical characteristics of the sample.

The results obtained for the figures of merit of the PLSR models are presented in Table 2. It can be observed that the average prediction errors (represented by RMSEC and RMSEP) were lower than 3.0% (m/m). For the samples presenting the lowest cocaine concentrations relative errors of approximately $\pm 20\%$ were observed. However, considering all validation samples the average relative error

Table 1. Results for the number of outliers identified by each parameter and the variation of the RMSEC and RMSEP values observed

Model ^a	Sample	No. of outliers detected in each test				RMSEC	RMSEP
		Leverage	X residuals	Y residuals	Total		
Reflectance M1	184	10	4	5	16	5.26	5.34
Reflectance M2	168	5	1	4	10	3.80	5.32
Reflectance M3	158	4 ^b	0	3	3	3.28	5.23
Reflectance M4 Opt	155	0	0	0	0	2.89	5.21
Reflectance _{Val}	91	1	0	6	7	2.89	5.21
Reflectance _{Val} Opt	84	0	0	0	0	2.89	2.47
Absorbance M1	184	10	4	6	18	5.52	5.26
Absorbance M2	166	4	1	2	6	3.67	5.29
Absorbance M3	160	2 ^b	0	2	2	3.10	5.21
Absorbance M4 Opt	158	0	0	0	0	2.70	5.16
Absorbance _{Val}	91	0	0	6	6	2.70	5.16
Absorbance _{Val} Opt	85	0	0	0	0	2.70	2.77

^aM: model; Opt: optimized; Val: validation set; ^bsamples not excluded.

was approximately 9% and 10% for the reflectance and absorbance models, respectively.

Table 2. Analytical figures of merit for PLS models for the properties of interest

Figure of merit		Reflectance	Absorbance
Trueness / % (m/m)	RMSEC	2.89	2.70
	RMSEP	2.47	2.77
	REP _{Cal}	3.94	3.66
	REP _{Val}	9.11	9.94
Precision / % (m/m)	1.46	1.52	
Sensitivity / % (m/m) ⁻¹	0.195	0.137	
Goodness of fit ^a	slope	0.89 ± 0.11	0.82 ± 0.11
	intercept	8.9 ± 9.1	14.6 ± 9.1
	corr coef (R)	0.904	0.914
MDC / % (m/m)		12.8	11.6

^a99% confidence interval.

The linearity of the method was evaluated by the distributions and histograms of the residuas of the PLSR models, which are presented in Figure 3. Visually it is possible to verify the random behavior of these distributions. However, to verify the assumption of independence and normality of the residuals it was applied the Jarque-Bera test.³² According to this test, with 95% of confidence, the normality of the discriptions cannot be rejected. Since the residual plot indicates the validity of the linear model, the

fitting of a straight line relating reference *versus* estimated values can be used to estimate a correlation coefficient, slope and intercept. These parameters may then be used to express the goodness of fit of the PLSR models.

Figure 4 presents the dispersion graphics of the regressions between the reference and estimated values for both PLSR models. Good agreement was observed between the evaluated methods (GC-FID and FTIR), with correlation coefficients higher than 0.90 for both models. However, the results for the slope and the intercept of the regression line between the reference and estimated values for the cocaine concentration (presented in Table 2) shows that the PLSR model developed with the absorbance data present both constant and proportional systematic errors. This can be seen by the fact that the confidence intervals (with 95% confidence) do not contain the expected values of 1 and 0 for the slope and intercept, respectively. On the other hand, the model developed with the reflectance data showed no significant systematic errors. Therefore, both models showed comparable prediction errors, but taking into account the goodness of fit, only the PLSR model developed with reflectance values may be considered to have adequate trueness. This was an unexpected result since according the Beer-Lambert law, the data in the absorbance scale should provide a better relation with the analyte concentration. However, it should be noted that the Beer-Lambert law is strictly valid only for transmittance measurements.

The uncertainty of the PLSR models was estimated in accordance with Pierna *et al.*³⁰ Considering this approach,

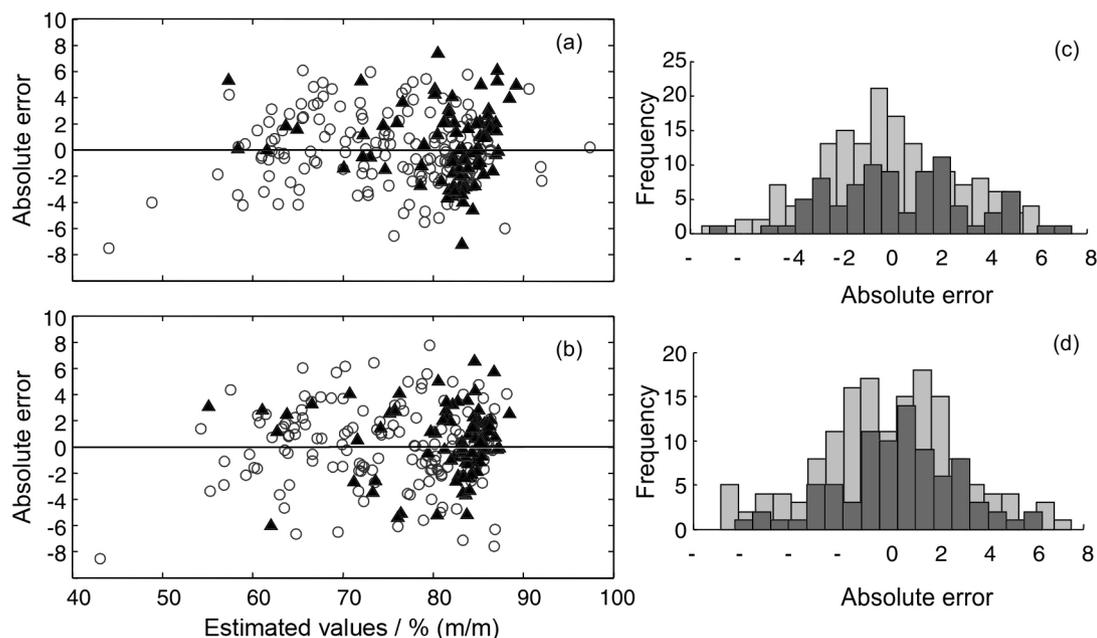


Figure 3. Distributions of the absolute errors obtained with the PLSR model developed with the absorbance (a) and reflectance data (b) against the estimated cocaine content values. Calibration (O) and validation samples (▲). Histograms of the absolute errors obtained with the absorbance (c) and reflectance (d) PLSR models (calibration data (light grey bars) and validation data (dark grey bars)).

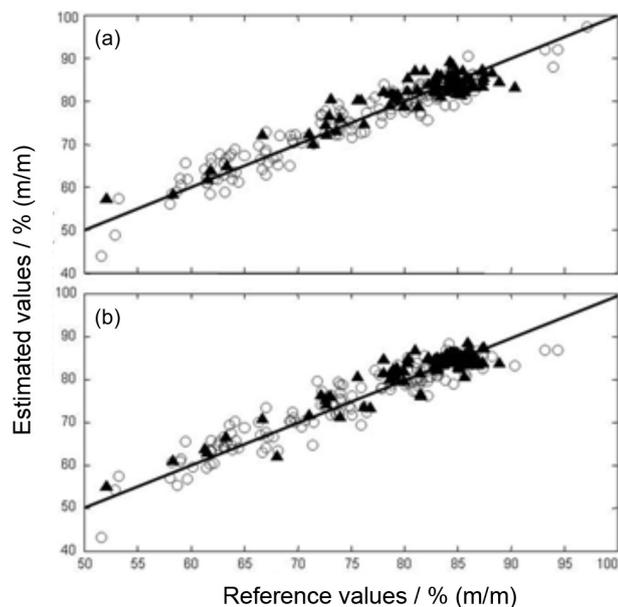


Figure 4. Reference values versus estimated values by the PLSR models for absorbance (a) and reflectance (b). Calibration samples (○) and validation samples (▲).

it was found that, for both models, the average uncertainty was approximately 7%, with 95% confidence. A plot with confidence intervals for some validation samples is shown in Figure 5, which illustrates the uncertainty of the results of the PLSR model developed with the reflectance data, and 95% confidence.

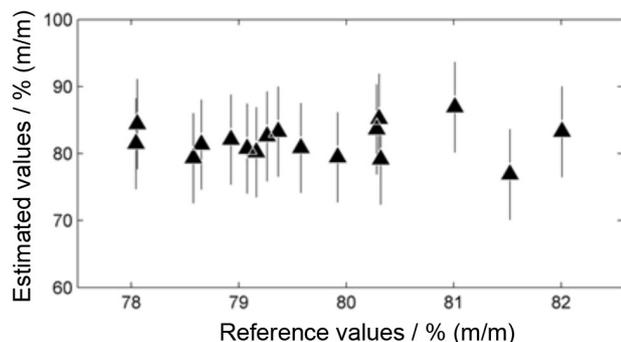


Figure 5. Confidence intervals for some validation samples, with 95% of confidence, obtained for the PLSR model developed with the reflectance data.

Precision at the repeatability level showed good results for both PLSR models, being approximately equal to 1.5% (m/m).

The MDC estimates represent an important figure of merit for the method. The estimated MDC values for the reflectance and absorbance models were 12.8% and 11.6%, respectively, which indicates that the FTIR method is suitable for determining the concentrations of most seizures made by BFP.

Conclusions

The results show that the validated method based on the combination of the ATR-FTIR spectroscopic technique and PLSR allows for the direct determination of the hydrochloride cocaine concentration of drug samples seized in several Brazilian states.

The method presented low absolute and relative average errors (lower than 3% (m/m) and 10%, respectively). Based on the quality of the fit, the model developed with the reflectance data was selected as the best model for determining cocaine hydrochloride concentration in drug samples.

This method can be considered convenient and versatile since it has the ability to significantly reduce the time and cost of analysis with respect to chromatographic analysis. Additionally, it is more environmentally friendly as it does not generate any chemical residues. Although gas chromatography has been frequently used because it offers accurate results, infrared spectroscopy has shown similar precision, acceptable trueness and detection capability, and a wide linear range (35% to 99% (m/m)), which fulfills the requirements for its application in forensic laboratories.

The proposed method allows for fast and accurate creation of criminal expert reports, thus contributing to the judiciary system and benefitting society.

Acknowledgments

The authors thank FINEP/MCT for the financial support of the PeQui Project (01.09.0275-00), CNPq, BRA 190/ UNODC/FP, INCTAA/CNPq and INCTBio/CNPq.

References

1. United Nations Office on Drugs and Crime (UNODC); *World Drug Report*; United Nations: New York, 2011.
2. United Nations Office on Drugs and Crime (UNODC); *World Drug Report*; United Nations: New York, 2013.
3. Chasin, A. A. M.; Mídio, A. F.; *Rev. Bras. de Toxicol.* **1991**, *4*, 23.
4. Moore, J. M.; Casale, J. F.; *J. Chromatogr. A* **1994**, *674*, 165.
5. Floriani, G.; Gasparetto, J. C.; Pontarolo, R.; Gonçalves, A. G.; *Forensic Sci. Int.* **2014**, *235*, 32.
6. Buján, L.; Fernández, P.; Lafuente, N.; Aldonza, M.; Bermejo, A. M.; *Anal. Chem. Lett.* **2001**, *34*, 2263.
7. Piñero, E. L.; Casale, J. F.; *Microgram J.* **2006**, *4*, 47.
8. United Nations Office on Drugs and Crime (UNODC); *Methods for Impurity Profiling of Heroin and Cocaine*, United Nations: New York, 2005.

9. Rodrigues, N. V. S.; Cardoso, E. M.; Andrade, M. V. O.; Donnici, C. L.; Sena, M. M.; *J. Braz. Chem. Soc.* **2013**, *24*, 507.
10. Ravreby, M.; *J. Forensic Sci.* **1987**, *32*, 20.
11. Ryder, A. G.; O'Connor, G. M.; Glynn, T. J.; *J. Forensic Sci.* **1999**, *44*, 1013.
12. Maharaj, R.; *J. of Third World Med.* **2009**, *7*.
13. Pérez-Alfonso, C.; Galipienso, N.; Garrigues, S.; Guardia, M.; *Forensic Sci. Int.* **2014**, *237*, 70.
14. Brereton, R. G.; *Chemometrics: Data Analysis for the Laboratory and Chemical Plant*, Wiley: Chichester, USA, 2003.
15. Martens, H.; Naes, T.; *Multivariate Calibration*, John Wiley & Sons: New York, USA, 1989.
16. Walczak, B.; Massart, D. L.; *Chemom. Intell. Lab. Syst.* **1998**, *41*, 1.
17. Kennard, R. W.; Stone, L. A.; *Technometrics* **1969**, *11*, 137.
18. Annual Book of ASTM Standards, *Standards Practices for Infrared, Multivariate, Quantitative Analysis*, ASTM International E1655-05, West Conshohocken: Pennsylvania, USA, 2005.
19. Valderrama, P.; Poppi, R. J.; Braga, J. W. B.; *J. Agric. Food Chem.* **2007**, *55*, 8331.
20. INMETRO, *Vocabulário Internacional de Metrologia- Conceitos Fundamentais e Gerais e Termos Associados* (VIM 2012), 1ª ed. luso-brasileira, Portaria No. 232, Brasília, Brasil, 2012.
21. Valderrama, P.; Braga, J. W. B.; Poppi, R. J.; *Quim. Nova* **2009**, *32*, 1278.
22. Lorber, A.; *Anal. Chem.* **1986**, *58*, 1167.
23. Olivieri, A. C.; Faber, M. N.; Ferré, J.; Boqué, R.; Kalivas, J. H.; Mark, H.; *Pure Appl. Chem.* **2006**, *78*, 633.
24. Instituto Nacional de Metrologia, Normalização e Qualidade Industrial (INMETRO); *Orientações sobre Validação de Métodos de Ensaio Químicos, DOQ-CGCRE-008*, Rio de Janeiro: Brasil, 2007.
25. Ministério da Agricultura, Pecuária e Abastecimento (MAPA). *Guia de Validação e Controle de Qualidade Analítica: Fármacos em Produtos para Alimentação e Medicamentos Veterinários*, Brasília: Brasil, 2011.
26. O'Malley, A. J.; Deely, J. J.; *Aust. N. Z. J. Stat.* **2003**, *45*, 43.
27. ISO, 11843-2: *Capability of Detection, International Standards Organization*, Switzerland: Geneva, 2000.
28. Ortiz, M. C.; Sarabia, L. A.; Herrero, A.; Sánchez, M. S.; Sanz, M. B.; Rueda, D.; Giménez, M. E.; Meléndez, M. E.; *Chemom. Intell. Lab. Syst.* **2003**, *69*, 21.
29. Boque, R.; Larrechi, M. S.; Rius, F. X.; *Chemom. Intell. Lab. Syst.* **1999**, *45*, 397.
30. Pierna, J. A. F.; Jin, L.; Wahl, F.; Faber, N. M.; Massart, D. L.; *Chemom. Intell. Lab. Syst.* **2003**, *65*, 281.
31. Van Der Voet, H.; *J. Chemom.* **1999**, *13*, 195.
32. Jarque, C. M.; Bera, A. K.; *Int. Stat. Rev.* **1987**, *55*, 163.

Submitted on: April 28, 2014

Published online: July 15, 2014