

Chlorination of Isatins with Trichloroisocyanuric Acid

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Synthesis and characterization of the chloroisatins

Typical procedure for the reaction in HOAc

To a suspension of TCCA and HOAc the isatin derivative was added. The reaction medium was kept stirring at different temperatures and times (Tables 1 and 2). The mixture was then poured over cracked ice. The precipitate was filtered under vacuum and washed thoroughly with water. Subsequently, the product was solubilized in ethyl acetate, filtered and the solution evaporated at reduced pressure to separate the isocyanuric acid formed as byproduct.

Typical procedure for the reaction in H_2SO_4

To a suspension of TCCA and H_2SO_4 was added the isatin derivative in an ice bath. After the complete addition of the isatin, the ice bath was removed, and the mixture was kept stirring during the time described in Table 3. The mixture was then poured over cracked ice. The precipitate was filtered under vacuum and washed thoroughly with water. Subsequently, the product was solubilized in ethyl acetate, filtered and the solution evaporated at reduced pressure to separate the isocyanuric acid formed as byproduct. The products (**9b**) and (**9c**) were separated by high performance liquid chromatography (HPLC) and analyzed separately by ¹H and ¹³C NMR.

Separation by HPLC

Separation by HPLC of (**9b**) and (**9c**) was performed on Agilent 1100 series chromatograph detection by ultraviolet (UV, λ_{max} /nm 280). Semi-preparative method in a reversed phase column C-18 (Agilent Zorbax Eclipse, 9.4 mm (diameter) × 250 mm (length), 5 micron) under isocratic conditions using 45% water and 55% methanol as mobile phase, flow 3 mL/min, with 30 min elution was employed.

Computational details

All calculations were carried out with the Gaussian 09 package¹ using the M06-2X functional with the 6-311++G** basis set. The optimized geometries were characterized as minima on the potential energy surface by the absence of imaginary vibrational frequencies. All energy differences correspond to enthalpies differences at 298K and 1 atm.

Spectroscopic data

1a: IR (film) ν_{max}/cm⁻¹: 3085, 2923, 1764, 1729, 1614, 1596, 1483, 1315, 1294, 1176, 1126, 950, 427. ¹H NMR δ (200 MHz, CDCl₃): 3.23 (3H, s), 6.85 (1H, d, *J* 8.0 Hz), 7.51-7.57 (2H, m); ¹³C NMR δ (50 MHz, CDCl₃): 26.5 (CH₃), 111.4 (CH), 118.3 (q), 125.2 (CH), 129.7 (q), 137.9 (CH), 149.8 (q), 157.8 (q), 182.4 (q); MS (70 eV) *m/z* 195 [M]⁺, 197 [M+2]⁺, 160, 104.

2a: IR (film) v_{max}/cm^{-1} : 3060, 1770, 1743, 1606, 1456, 1423, 1307, 1270, 1178, 1160, 827, 717, 466. ¹H NMR δ (200 MHz, CDCl₃): 7.07 (1H, d, *J* 8.0 Hz), 7.71 (1H, d, *J* 2.0 Hz), 7.81 (1H, dd, *J* 2.0 and 8.0 Hz); ¹³C NMR δ (50 MHz, CDCl₃): 113.0 (CH), 118.0 (q), 120.5 (q), 127.9 (CH), 141.2

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(CH), 148.0 (q), 154.7 (q), 176.8 (q), 182.4 (q); MS (70 eV) *m*/*z* 259 [M]⁺, 261, [M+2]⁺, 263 [M+3]⁺, 168, 226.

2b: IR (film) v_{max}/cm^{-1} : 3471, 3174, 3110, 1754, 1610, 1457, 1425, 1290, 1224, 1168, 837; ¹H NMR δ (200 MHz, CDCl₃+DMSO): 7.47 (1H, s), 7.58 (1H, s), 11.43 (1H, br s, NH); ¹³C NMR δ (50 MHz, CDCl₃+DMSO): 114.8 (q), 118.2 (q), 119.5 (q), 125.5 (CH), 139.1 (CH), 146.8 (q), 158.3 (q), 182.1 (q); MS (70 eV) *m*/*z* 259 [M]⁺, 261, [M+2]⁺, 263 [M+3]⁺, 233, 205.

5b: IR (film) v_{max}/cm^{-1} : 3467, 3423, 1751, 1727, 1608, 1457, 1467, 1326, 1176, 1108, 827; ¹H NMR δ (200 MHz, CDCl₃+DMSO): 3.16 (3H, s), 6.89 (1H, d, *J* 8.0 Hz), 7.43 (1H, s), 7.52 (1H, d, *J* 8.0 Hz), 11.43 (1H, br s, NH); ¹³C NMR δ (50 MHz, CDCl₃+DMSO): 25.4 (CH₃), 116.6 (CH), 123.1 (q), 129.5 (CH), 134.0 (q), 142.6 (CH), 154.6 (q), 162.6 (q), 187.3 (q); MS (70 eV) *m*/*z* 195 [M]⁺, 197 [M+2]⁺, 167, 139.

6b: IR (film) v_{max}/cm^{-1} : 3452, 3415, 3178, 3102, 1743, 1616, 1469, 1297, 889, 700; ¹H NMR δ (200 MHz, CDCl₃+DMSO): 2.15 (3H, s), 7.20 (1H, br s), 7.27 (1H, br s), 11.13 (1H, br s, NH); ¹³C NMR δ (50 MHz, CDCl₃+DMSO): 14.2 (CH₃), 117.0 (q), 120.3 (CH), 122.8 (q), 126.3 (CH), 137.2 (CH), 146.9 (q), 158.3 (q), 182.6 (q); MS (70 eV) *m*/*z* 195 [M]⁺, 197 [M+2]⁺, 167, 139, 104.

7b: IR (film) v_{max}/cm^{-1} : 3488, 3473, 3180, 3114, 1743, 1754, 1621, 1600, 1479, 1299, 1211, 1099, 925, 867, 796; ¹H NMR δ (200 MHz, CDCl₃+DMSO): 6.92 (1H, d *J* 7.0 Hz), 7.04 (1H, d *J* 7.0 Hz), 11.04 (1H, br s, NH); ¹³C NMR δ (50 MHz, CDCl₃+DMSO): 110.1 (CH d *J* 27 Hz), 117.7 (q, d *J* 7.0 Hz), 118.6 (q, d, *J* 7.0 Hz) 124.4 (CH, d, *J* 27 Hz), 144.3 (q, d, *J* 2.5 Hz), 157.7 (q, d, *J* 246.5 Hz), 158.9 (q), 182.8 (q, d *J* 2.5 Hz). MS (70 eV) *m/z* 199 [M]⁺, 201 [M+2]⁺, 171, 143.

9c: IR (film) v_{max}/cm^{-1} : 3193, 3182, 3099, 1770, 1745, 1610, 1450, 1164, 983, 688; ¹H NMR δ (200 MHz, CDCl₃+DMSO): 7.71 (1H, d *J* 1.5 Hz), 7.77 (1H, d *J* 1.5 Hz), 10.78 (1H, br s, NH); ¹³C NMR δ (50 MHz, CDCl₃+DMSO): 84.79 (q), 118.97 (q), 120.36 (q), 132.03 (CH), 145.34 (CH), 147.48 (q), 158.23 (q), 182.14 (q); MS (70 eV) *m/z* 307 [M]⁺, 309 [M+2]⁺, 279, 251.

11b: IR (film) v_{max} /cm⁻¹: 3479, 3465, 3268, 3087, 1766, 1791, 1594, 1558, 1388, 1247, 958; ¹H NMR δ (200 MHz, CDCl₃+DMSO): 6.29 (1H, s), 10.40 (1H, br s, NH); ¹³C NMR δ (50 MHz, CDCl₃+DMSO): 114.3 (CH), 118.7 (q), 126.3 (q), 127.8 (q), 129.5 (q), 148.6 (q), 156.4 (q), 178.5 (q); MS (70 eV) *m/z* 337 [M]⁺, 339 [M+2]⁺, 341 [M+3]⁺, 343 [M+4]⁺, 311, 284, 204.

¹H and ¹³C NMR spectra







Figure S2. ¹³C NMR of 1a.



Figure S3. ¹H NMR of 2a.



Figure S4. ¹³C NMR of 2a.



Figure S5. ¹H NMR of 2b.





Figure S6. ¹³C NMR of 2b.











Figure S9. ¹H NMR of 6b.



6b

Figure S10. ¹³C NMR of 6b.







Figure S11. ¹H NMR of 7b.



Figure S12. ¹³C NMR of 7b.



Figure S13. ¹H NMR of 9c.



Figure S14. ¹³C NMR of 9c.













Figure S17. FTIR spectra of 1a.



Figure S18. FTIR spectra of 2a.



Figure S19. FTIR spectra of 2b.



Figure S21. FTIR spectra of 6b.





Figure S22. FTIR spectra of 7b.



Figure S23. FTIR spectra of 9c.





Figure S24. FTIR spectra of 11b.

Mass Spectra



Figure S25. Mass spectrum of 1a.



Figure S26. Mass spectrum of 2a.



Figure S27. Mass spectrum of 2b.



Figure S28. Mass spectrum of 4b.



Figure S29. Mass spectrum of 5b.



Figure S30. Mass spectrum of 6b.



Figure S31. Mass spectrum of 7b.



Figure S32. Mass spectrum of 8b.



Figure S33. Mass spectrum of 8c.



Figure S34. Mass spectrum of 9b.



Figure S35. Mass spectrum of 9c.

Scan 647 (9.569 min): CLO46BP2.D



Figure S36. Mass spectrum of 11b.



Figure S37. Mass spectrum of 11c.