

## Determination of Tetracycline in Environmental Water Samples at a Graphite-Polyurethane Composite Electrode

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O eletrodo compósito grafite-poliuretana foi avaliado na determinação de tetraciclina (TC) em amostras de água natural. Utilizando voltametria de pulso diferencial (DPV), uma resposta linear foi observada no intervalo de 4,00-40,0  $\mu\text{mol L}^{-1}$  com limite de detecção de 2,80  $\mu\text{mol L}^{-1}$ , sem necessidade de renovação da superfície entre medidas sucessivas. Durante determinação de tetraciclina em amostras de água, foram obtidas recuperações entre 92,6 e 100%. Os resultados para determinação de TC em amostras de água após a etapa de pré-concentração concordam com valor adicionado em um nível de confiança de 95% de acordo com o teste *t-student*.

A bare graphite-polyurethane composite was evaluated in the tetracycline (TC) determination in natural water samples. Using differential pulse voltammetry (DPV), a linear response was observed in the range of 4.00-40.0  $\mu\text{mol L}^{-1}$  with limit of detection of 2.80  $\mu\text{mol L}^{-1}$ , without the need of surface renewing between successive runs. During the tetracycline determination in water samples, recoveries between 92.6 and 100% were found. The results for TC determination in water samples after a pre-concentration stage agreed with spiked value at a 95% confidence level according to student *t*-test.

**Keywords:** tetracycline, graphite-polyurethane composite electrode, natural water

### Introduction

Tetracyclines (TCs, Figure 1) are a family of antibiotics that have been used for more than 50 years in the treatment of bacterial infections in both humans and animals.<sup>1</sup>

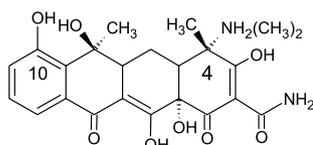


Figure 1. Chemical structure of tetracycline.

Tetracycline residues in biological samples can be directly toxic or cause allergic reactions in some hypersensitive individuals, also promoting the occurrence of antimicrobial-resistant bacteria. However, its concentration is still very low in that samples.<sup>2</sup> To prevent health problems, some countries from the European Union and the United States have established their own maximum residue limits for these compounds in livestock products.<sup>3-5</sup>

Tetracyclines are also active against a wide range of *gram*-(+) and *gram*-(-) aerobic and anaerobic bacteria, including *Spirochete*, *Actinomyces*, *Rickettsia* and *Mycoplasma*.<sup>6</sup>

Even though tetracyclines may be considered carcinogenic<sup>7,8</sup> and have been implicated in the growing prevalence of antibiotic resistance in humans,<sup>9,10</sup> they are still largely used against infections, such as respiratory tract infections, urethritis and severe acne. It also has a role in the treatment of multidrug resistant malaria. Adverse effects include gastrointestinal disturbances, renal dysfunction, hepatotoxicity, raised intracranial pressure and skin infections, like rosacea and perioral dermatitis.<sup>11</sup>

These antibiotics also have non-therapeutic use since they are widely used in veterinary practice as food additives for cattle, pigs, sheep, poultry and fish. In these cases, they are used in sub-therapeutic doses for growth promotion due to their broad range of activity against bacteria and low cost.<sup>12,13</sup> So, there is a risk that residues of tetracycline or its metabolites are present in foodstuffs from animals treated with them. This represents an impact on human health because the TCs strongly adsorb onto

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environmental materials in which they also maintain their chemical activity.<sup>14,15</sup>

Several methods have been used for the tetracycline determination, including capillary electrophoresis,<sup>5,16</sup> fluorescence,<sup>14</sup> UV-Vis spectroscopy,<sup>17</sup> spectrofluorimetric,<sup>18</sup> liquid chromatography-tandem mass spectrometry<sup>19,20</sup> and high performance liquid chromatography.<sup>6,21</sup>

Although chromatographic procedures usually offer lower limits of detection, electrochemical methods can be an interesting alternative due to their simplicity, less analysis time, low cost and small amount of waste generation.

Voltammetric determination of tetracycline was performed using a disposable screen-printed gold electrode,<sup>12</sup> a multi-walled carbon nanotube-ionic liquid film coated glassy carbon electrode<sup>22</sup> and an anodized boron-doped diamond thin film electrode.<sup>23</sup> In these works, the limits of detection  $0.96 \mu\text{mol L}^{-1}$ ,<sup>12</sup>  $0.3 \text{ nmol L}^{-1}$ <sup>22</sup> and  $10 \text{ nmol L}^{-1}$ <sup>23</sup> were measured.

A flow injection analysis of TC with pulsed amperometric detection was also developed, with sensitivity of  $13.7 \mu\text{A L mmol}^{-1}$ .<sup>24</sup> Molecularly imprinted polymers (MIPs) were developed by precipitation polymerization using TCs as template.<sup>25,26</sup> O'Connor and Aga<sup>27</sup> described the analysis of tetracycline antibiotics in soil and Seifrtová *et al.*<sup>28</sup> presented analytical methodologies for the determination of these antibiotics in environmental waters. The stability of tetracyclines in water was discussed by Loftin *et al.*<sup>29</sup> According to these authors, tetracyclines undergo degradation in water, including hydrolysis and epimerization. The effects of ionic strength, pH and temperature on such reactions were reported, and for tetracycline at  $22 \text{ }^\circ\text{C}$  in  $4 \text{ mmol L}^{-1}$  phosphate buffer pH 2, a  $t_{1/2}$  of 306 h was found.<sup>29</sup>

The purpose of this work is to investigate the voltammetric behavior and the possibility of TC determination using a 60% (graphite, m/m) bare graphite-polyurethane composite (GPU) electrode in environmental water samples. This composite was proposed<sup>30</sup> and showed to be sensitive and useful in differential pulse voltammetry (DPV),<sup>31</sup> square wave voltammetry (SWV)<sup>32</sup> and as an amperometric detector in flow injection analysis<sup>33</sup> of antihypertensives.

## Experimental

### Apparatus

Voltammetric experiments were performed using an  $\mu\text{AUTOLAB}$  Type III potentiostat/galvanostat (Ecochemie, The Netherlands) coupled to a personal computer and controlled with a GPES 4.9 software (Ecochemie, The Netherlands). A homemade three-electrode glass cell with 25.0 mL full capacity was used with a saturated calomel

electrode (SCE) and a platinum wire as the reference and counter electrodes, respectively. The 60% (graphite, m/m) GPU<sup>30</sup> was used as a working electrode. All measurements were performed at room temperature without need of degassing the solution.

### Reagents and solutions

All reagents used were of the analytical grade and used as received. Solutions were prepared with water purified in a Barnstead™ EasyPure® RoDi (Thermoscientific, D13321, resistivity  $\geq 18 \text{ M}\Omega \text{ cm}$ ) system. Tetracycline hydrochloride (95%) was purchased from Sigma-Aldrich (USA), and a  $2.00 \times 10^{-3} \text{ mol L}^{-1}$  TC stock solution in  $0.1 \text{ mol L}^{-1}$  phosphate buffer pH 2.3 was freshly prepared (just before the measurements) and used for no more than 12 h. Considering the findings of Loftin *et al.*,<sup>29</sup> the degradation is negligible under such conditions.

### Preparation of the composite electrode

The composite used as working electrode was prepared as previously described in literature.<sup>30</sup> Briefly, following the manufacturer instructions, polyurethane (PU, from Poliquil, Araraquara, Brazil) was prepared by taking equal mass amounts of the polyol and hardener, to this mixture, an appropriate amount of graphite (Aldrich, 1-2  $\mu\text{m}$ , USA) was added. These components were mixed in a mortar for 10 min in order to obtain a homogeneous mixture containing 60% of graphite (m/m).

The resulting mixture was inserted in a casting mould and extruded as 3.0 mm diameter rods with a manual press. The composite was cured for 24 h at room temperature and cut in 1.0 cm long rods. A copper wire ( $\phi = 1.0 \text{ mm}$ ) was attached to these rods by a silver epoxy (EPO-TEK 410E, Epoxy Technology, USA) (a conducting resin that establishes the electric contact). This set was then inserted in a 5.0 mm i.d. glass tube and sealed with epoxy resin (Silaex, SQ 2004, Brazil). The electrodes were allowed to cure for additional 24 h.

After curing, the resin in excess was removed using a 600-grit abrasive paper, and the electrode surface polished with  $1.0 \mu\text{m}$   $\gamma\text{-Al}_2\text{O}_3$  suspension in an APL-2 polishing wheel (Arotec, Brazil), and sonicated in isopropyl alcohol and water for 5 min in each solvent.

### Procedures

Voltammetric measurements were performed using the 60% (graphite, m/m) GPU composite ( $\phi = 3.0 \text{ mm}$  diameter) as working electrode.

In order to record all voltammograms, appropriate aliquots of a freshly prepared TC stock solution were diluted in 20.0 mL of phosphate buffer pH 2.3 inside the voltammetric cell.

Pulse amplitude ( $a$ ) and scan rate ( $v$ ) were optimized for the DPV experiments from a set of values:  $a = 10$  and  $50$  mV, and  $v = 10$  and  $50$  mV s<sup>-1</sup>, according to a 2<sup>n</sup> factorial planning, resulting in four experiments, in phosphate buffer pH 2.3.

#### Procedures for environmental water sample analysis<sup>34</sup>

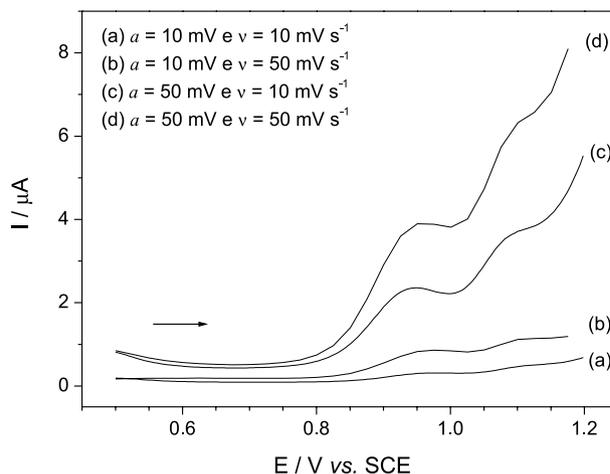
Natural water sample was collected from a lake located in São Carlos City (São Paulo State, Brazil) at 21°59'08"-S and 47°52'58"-W. This collect was done using amber glass bottles that were pre-cleaned with HPLC grade methanol and rinsed with ultrapure water before use. Each water sample was filtered firstly using a filter paper, and then glass fiber membrane filter (0.45 μm) (Sartorius, Germany) to remove solid particulate matter. The natural water sample (500 mL) was then acidified to pH 3.0 with 3.0 mmol L<sup>-1</sup> H<sub>2</sub>SO<sub>4</sub>. An aliquot of 100 μL of 1.0 × 10<sup>-2</sup> mol L<sup>-1</sup> TC was spiked to 500 mL of this natural water sample, resulting in a water sample containing 2.00 μmol L<sup>-1</sup> of TC.

The solid phase extraction (SPE) procedure was performed using Oasis<sup>®</sup> HLB 30 μm Extraction Cartridge (200 mg, 6.0 mL) (Waters, USA). The cartridges were preconditioned with 6.0 mL of methanol (HPLC grade) and 6.0 mL of ultrapure water, at a flow rate of 2.0 mL min<sup>-1</sup>.

After the conditioning step, the spiked water samples were percolated through the cartridges at a flow rate of 10.0 mL min<sup>-1</sup>. Elution was performed with 3 × 2.0 mL of methanol at 2.0 mL min<sup>-1</sup>. The extract was evaporated under a gentle nitrogen stream in a volumetric flask and reconstituted with 100.0 mL of phosphate buffer pH 2.3 to obtain a solution corresponding to 10.0 μmol L<sup>-1</sup> in TC. Then, additions of adequate volumes of 2.00 mmol L<sup>-1</sup> TC standard solution were made, and differential pulse voltammograms were recorded in triplicate, as the procedure was repeated three times. This procedure was also repeated in triplicate with TC solutions of ultrapure water to evaluate the efficiency of the extraction procedure.

## Results and Discussion

DPV voltammograms of 1.00 × 10<sup>-4</sup> mol L<sup>-1</sup> TC solutions in phosphate buffer pH 2.3 are presented in Figure 2, using 10 and 50 mV pulse amplitude at 10 and 50 mV s<sup>-1</sup> scan rates based on a 2<sup>n</sup> factorial planning. In these voltammograms, two oxidation peaks appeared at



**Figure 2.** Differential pulse voltammograms obtained at 60% (graphite, m/m) GPU composite electrode using 1.00 × 10<sup>-4</sup> mol L<sup>-1</sup> TC in phosphate buffer pH 2.3.

ca. 960 and 1100 mV (vs. SCE) at the GPU composite electrode.

Better peak definition was obtained using  $a = 50$  mV, while higher sensitivities were reached with  $v = 10$  mV s<sup>-1</sup>. For  $a = 10$  mV, very low sensitivity was achieved. Thus, 50 mV and 10 mV s<sup>-1</sup> were used as pulse amplitude and scan rate, respectively, in further studies since less distortion on the peak shapes were observed under such conditions, as can be observed in Figure 2.

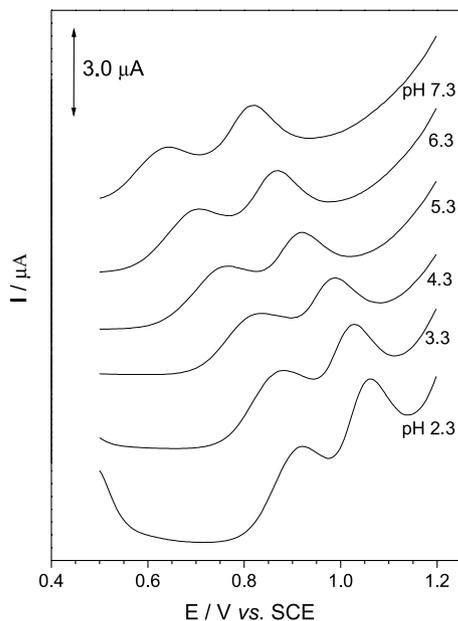
Although apparently higher currents were observed in Figure 2 curve d, there is a very high residual current on these voltammograms making the actual faradaic current not as higher as than in curve c.

#### Effect of pH on the DPV voltammograms

Figure 3 depicts the differential pulse voltammograms of TC in phosphate buffer pH 2.3 to 7.3 at the 60% (graphite, m/m) GPU composite electrode. As presented by these results, the oxidation currents decreased with pH increasing over the whole range investigated, while the peaks shifted to less positive potentials. Therefore, it is evident that the two peaks around 920 and 1060 mV (vs. SCE) are better resolved in pH 2.3.

Vega *et al.*<sup>15</sup> described only one oxidation peak at a multi-walled carbon nanotube modified glassy carbon electrode and attributed the signal decreasing at higher pH to the low stability of TC in these conditions.

Wangfuengkanagul *et al.*<sup>23</sup> using a diamond electrode doped with boron observed the highest current signal for TC at pH 2. They attributed this to the fact that the electrode is negatively charged, while the TC would be a cation at that pH.<sup>23</sup> However, this does not seem to be the only cause for



**Figure 3.** Differential pulse voltammograms obtained at 60% (graphite, m/m) GPU composite electrode using  $1.00 \times 10^{-4}$  mol L<sup>-1</sup> TC solution in phosphate buffer, with different pH values (2.3-7.3), with  $a = 50$  mV and  $v = 10$  mV s<sup>-1</sup>.

the best response in an acidic medium since in this work it was observed the same fact, without the negative charges on the electrode surface.

Masawat and Slater<sup>12</sup> also observed best voltammetric responses and good resolution for both peaks at a gold electrode in acidic medium. Chatten *et al.*<sup>35</sup> suggest the dimethylamino group or phenol as likely sites for the initial oxidation at a potential less than +1.0 V (*vs.* SCE). The amino groups are protonated in the two aqueous acetate buffers pH 4.0 and 5.6 used by those authors, leading them to conclude that the phenolic moiety of the molecule undergoes an one electron irreversible oxidation.<sup>35</sup>

On the other hand, Dang *et al.*<sup>36</sup> observed only one TC oxidation peak around +0.58 V (*vs.* SCE) and attributed this peak to the phenolic substituent in position C-10 of TC. The mechanism of electrode process proposed by these authors was based on the oxidation of phenol group to benzoquinone, which is adsorbed at the acetylene black electrode surface.<sup>36</sup>

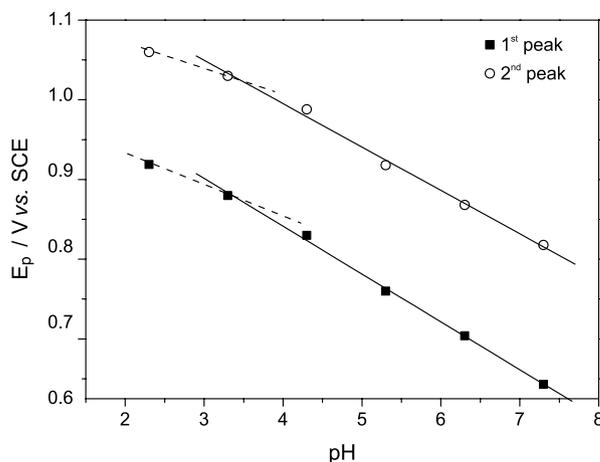
From the peak potential ( $E_p$ ) *vs.* pH plots (Figure 4), it can be observed a break around pH 3.5, with a well-defined linear region between pH 4.3 and 7.3. This break is visible for both TC oxidation peaks and evidences that the pKa is near this region. A more careful analysis of such breaks suggests 3.7 and 3.6 as the approximated values of pKa for the first and second TC oxidation peaks, respectively. Both are close to the value described in literature (pKa of 3.3).<sup>37</sup>

A linear dependence of peak potential between pH 4.3 and 7.3 obeyed the following relationships:

$$\begin{aligned} 1^{\text{st}} \text{ peak: } E_p &= 1081 \text{ mV} - 60 \text{ pH} \\ (n = 5 \text{ and correlation coefficient of } 0.999) \end{aligned} \quad (1)$$

$$\begin{aligned} 2^{\text{nd}} \text{ peak: } E_p &= 1213 \text{ mV} - 54 \text{ pH} \\ (n = 5 \text{ and correlation coefficient of } 0.997) \end{aligned} \quad (2)$$

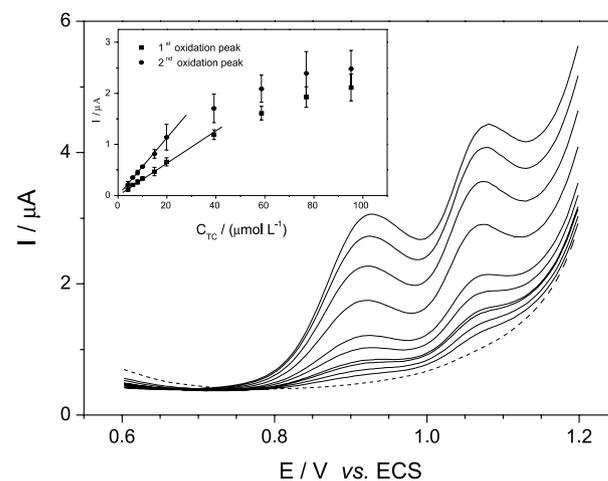
The slopes of these equations (pH 4.3-7.3) are close to 59 mV suggesting that the same number of protons and electrons are involved in the redox process, in these cases.



**Figure 4.** Relationship between the peak potential in differential pulse voltammetry and pH, in phosphate buffer solution at different pH values, containing  $1.00 \times 10^{-5}$  mol L<sup>-1</sup> TC.

#### Analytical curve

Using these optimized conditions, an analytical curve was obtained in the range of 4.00 to 95.0  $\mu\text{mol L}^{-1}$  TC. TC concentrations lower than this range did not present well-defined current signals. The curves in Figure 5



**Figure 5.** Dependence of the Differential pulse voltammograms with TC concentration from 4.00 to 95.0  $\mu\text{mol L}^{-1}$  in phosphate buffer, pH = 2.3,  $a = 50$  mV,  $v = 10$  mV s<sup>-1</sup> at the 60% (graphite, m/m) GPU composite electrode. The analytical curve is presented in the insert.

were obtained with no need for surface renewal between successive determinations, suggesting that there is no adsorption of the analyte or its oxidation products onto the electrode surface.

The DPV voltammograms showed successive current increase of both peaks when the TC concentration increases. To the first peak, a linear region was determined between 4.00 to 40.0  $\mu\text{mol L}^{-1}$  in the investigated interval, obeying the following linear equation:

$$I = 0.01 \mu\text{A} + 0.03 \mu\text{A} \mu\text{mol}^{-1} \text{L} C_{\text{TC}},$$

(n = 7, correlation coefficient of 0.996) (3)

The limit of detection determined as three times the standard deviation of the blank ( $S_d$ ) divided by the angular coefficient of straight line (b)<sup>38</sup> was 2.30  $\mu\text{mol L}^{-1}$ . To the second peak, the peak currents in the interval between 4.00 to 20.0  $\mu\text{mol L}^{-1}$  obeyed the linear equation:

$$I = 0.01 \mu\text{A} + 0.06 \mu\text{A} \mu\text{mol}^{-1} \text{L} C_{\text{TC}},$$

(n = 6, correlation coefficient of 0.998) (4)

In this case, the limit of detection was 2.60  $\mu\text{mol L}^{-1}$ .

#### Environmental water sample analysis

The 60% (graphite, m/m) GPU composite electrode was used in the analysis of environmental water that was spiked to be 2.00  $\mu\text{mol L}^{-1}$  TC. This spiking level corresponds to one that may be determined considering the linear range of the proposed procedure.

The sample treatment described in the Experimental section was adapted from the one presented by Gros *et al.*,<sup>39</sup> that is a recent method developed for the determination of pharmaceuticals in surface and wastewater.

The standard addition method was used for the voltammetric determination of the TC amount present in the water samples. The results obtained for the analysis of three different samples of environmental water containing a spiked amount of 2.00  $\mu\text{mol L}^{-1}$  resulted in  $1.93 \pm 0.07 \mu\text{mol L}^{-1}$ , which represents a recovery of  $96 \pm 3\%$ .

Moreover, a blank sample prepared with water purified in a Barnstead<sup>TM</sup> system and spiked with the analyte was also analyzed to test the efficiency of the extraction procedure. The results pointed to a recovery of  $98 \pm 2\%$ , for a 2.00  $\mu\text{mol L}^{-1}$  TC solution.

The results agreed at a confidence level of 95% according to student *t*-test, thus demonstrating the suitability of the method for determining these levels of tetracycline in water samples with the help of a pre-concentration step.

A single electrode was used during all measurements here described, showing that the composite electrode has a long useful life. The results obtained in the determination of tetracycline in natural water samples point to even lower limits of detection if the extraction procedure is optimized.

## Conclusions

The unmodified composite 60% GPU (graphite, m/m) electrode represents an interesting alternative to be used in differential pulse voltammetry for the tetracycline determination with limit of detection in the  $\mu\text{mol L}^{-1}$  level. Although even lower limit of detection were described in literature,<sup>12,22,23</sup> the method here proposed uses a non-modified and low cost electrode, when compared with gold,<sup>12</sup> carbon-nanotube-ionic liquid film<sup>22</sup> and non-commercial boron-doped diamond thin film.<sup>23</sup> In addition, the GPU electrode is more physically resistant than films. With the results here described (as a first approach), it can be inferred that using other pre-concentration strategies, even sub- $\mu\text{mol L}^{-1}$  concentrations can be detected.

The method was satisfactorily applied for TC determination in purified water as well as in lake water samples, revealing no interference from other interferents in the natural environment. Although the higher limits of detection, when compared with tandem mass-chromatography coupled procedures, the method can be considered as a screening procedure with much lower instrumental and analysis costs, and much lower amount of waste generation.

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