The preparation, characterization, theoretical calculations and biological application of four Ru(II) complexes with 2-picolinate (pic), 2,2’-bipyridine (bipy) and P-P as ligands \([P-P=1,1\text{-bis(diphenylphosphino)methane (dppm-1)}, 1,2\text{-bis(diphenylphosphino)ethane (dppe-2)}, 1,3\text{-bis(diphenylphosphino)propane (dppp-3)} \text{ or } 1,1’\text{-bis(diphenylphosphino)ferrocene (dppf-4)}]\), is here presented. The complexes 1-4, with general formula [Ru(pic)(P-P)(bipy)]PF_6, were characterized by elemental analysis and by infrared (IR), UV-Vis, nuclear magnetic resonance (NMR 1H and 13P{1H}) spectroscopies, cyclic voltammetry and X-ray crystallography technique. Additionally, preliminary in vitro tests against human breast (MDA-MB-231) and murine ascitic sarcoma 180 (S180) tumor cell lines were carried out, and compared with cisplatin, a reference drug. The drug concentration at which 50% of the cells are viable relative to the control (IC\(_{50}\)) values found for complexes 1, 2, 3 and 4 against MDA-MB-231 tumor cells were around 14.6, 7.6, 3.3 and 0.4 μM, respectively, while against S180 tumor cells these complexes showed IC\(_{50}\) values of 71.9, 31.3, 11.2 and 3.5 μM, respectively. Therefore, the complexes were more active against MDA-MB-231 than S180.

**Keywords:** ruthenium(II), picolinate, biphosphines, murine ascitic sarcoma 180, human breast cancer

**Introduction**

Metal based therapeutics are a precious class of drugs in cancer treatment, and ruthenium-based complexes are promising alternative candidates to antineoplastic therapeutics, because this class of compounds have shown selective bioactivity and the good ability to overcome the resistance that platinum-based therapeutics face. Another advantage to use ruthenium complexes as drug is that in some cases it has been shown these kind of compounds containing organic drugs, such as ligands, can overcome resistance developed by bacteria to the organic compounds alone. Recently, a great number of ruthenium complexes containing different ligands, have been tested as antitumor agents against different kinds of tumor cell lines, showing to be very active. Thus, picolinate containing complex [Ru(η⁶-p-cymene)Cl(picolinate)] is active against HeLa (drug concentration at which 50% of the cells are viable relative to the control (IC\(_{50}\)) = 82.0 μM) and human melanoma cells (FemX, IC\(_{50}\) = 36.2 μM). The possible reason for this behavior is that the [Ru(η⁶-p-cymene)Cl(picolinate)] accumulates in the deoxyribonucleic acid (DNA) molecule, since it has high affinity for DNA-binding.
According to Gligorijević et al., a possible reason for this is the obstruction of complex/DNA interactions. The rotation of the ligand around the Ru–N bond in complexes \([\text{Ru}(\eta^6-p\text{-cymene})(L)_2]\Cl_2 (L: \text{3-acetylpyridine, 2-amino-5-chloropyridine}) and the presence of –COOH in \([\text{Ru}(\eta^6-p\text{-cymene})(HL)\text{Cl}]) (HL: 2,3-pyridinedicarboxylic acid, 2,4-pyridinedicarboxylic acid) can impair the complex intercalation through the \(\eta^6\)-arene group.\[11\]

In particular, phosphine ligands can form complexes with various d-block metals (Au, Ag, Cu, Ru, Pd, Pt) and some of these compounds have been evaluated as potential antitumor agents against human tumor cell lines.\[12-17\] In addition, the coordination of picolinate ligand together with some phosphines, in metal complexes, might enhance their biological activity.\[18,19\]

Preliminary results obtained by our research group for ruthenium(II) complexes containing 2,2′-bipyridine (bipy) and its derivatives, as well as diphosphines as ligands, are very promising as anticancer agents.\[20-22\] These results have encouraged us to explore these kinds of systems, not only because of the good results obtained, but also because it is possible to investigate the relationship between the structural parameter of the complexes and their biological activity.\[8\]

In vitro cytotoxicity assays on cultured human tumor cell lines still represent the standard method for the initial screening of potential antitumor agents. Thus, as a first step in assessing their pharmacological properties, the new ruthenium complexes were assayed against human breast tumor cell lines MDA-MB-231 and murine ascitic sarcoma 180 tumor cells (S180) (ATCC TIB-66). The cells were routinely maintained in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% fetal bovine serum (FBS), at 37°C in a humidified 5% CO2 atmosphere. After reaching confluence, the cells were detached by trypsinization and counted. For the cytotoxicity assay, 5 × 10^4 cells well−1 were seeded in 200 μL of complete medium in 96-well assay microplates (Corning Costar). The plates were incubated at 37°C in 5% CO2 for 24 h to allow cell adhesion, prior to drug testing. All tested compounds were dissolved in sterile dimethyl sulfoxide (DMSO, stock solution with maximum concentration of 20 mmol L−1) and diluted to 5.0, 2.0, 1.0, 0.5, 0.2, 0.02 and 0.002 mmol L−1. From each of these dilute samples, 2 μL aliquots were added to 200 μL medium (without FBS) giving a final concentration of DMSO of approximately 1% and a final concentration of the complex diluted about 100 times. Attached cells were exposed to the compounds for a 24 h period. Cell respiration, as an indicator of cell viability, was then determined by the mitochondrial-dependent reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) to formazan.\[23\] MTT solution (0.5 mg mL−1) was added to cell cultures and incubated for 3 h, after which the medium was removed and cell were washed one time with Phosphate buffered saline (PBS). After washing, 100 μL of DMSO was added and the plates were shaken for 10 min to facilitate the solubilization of formazan. The plates were read at 570 nm (absorbance) using a microplate reader (Techmate 116, Bio-Tek Instruments). The IC50 value was calculated from a graph of absorbance vs concentration using Graphpad prism software. The experimental procedure used in this work was previously described.\[26-29\] The analysis of the molecular orbital (MO) compositions in terms of fragment MO’s (Frontier Orbital-FO) were performed using AOMix-CDA.\[30\] The molecular orbitals were visualized using Chemissian software.\[31\]

Partition coefficient (P)

Water-octanol partition coefficients were determined using the stir-n-octanol method, as previously described.\[8\]

In vitro cytotoxicity

In vitro cytotoxicity assays on cultured human tumor cell lines still represent the standard method for the initial screening of potential antitumor agents. Thus, as a first step in assessing their pharmacological properties, the new ruthenium complexes were assayed against human breast tumor cell lines MDA-MB-231 and murine ascitic sarcoma 180 tumor cells (S180) (ATCC TIB-66). The cells were routinely maintained in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% fetal bovine serum (FBS), at 37°C in a humidified 5% CO2 atmosphere. After reaching confluence, the cells were detached by trypsinization and counted. For the cytotoxicity assay, 5 × 10^4 cells well−1 were seeded in 200 μL of complete medium in 96-well assay microplates (Corning Costar). The plates were incubated at 37°C in 5% CO2 for 24 h to allow cell adhesion, prior to drug testing. All tested compounds were dissolved in sterile dimethyl sulfoxide (DMSO, stock solution with maximum concentration of 20 mmol L−1) and diluted to 5.0, 2.0, 1.0, 0.5, 0.2, 0.02 and 0.002 mmol L−1. From each of these dilute samples, 2 μL aliquots were added to 200 μL medium (without FBS) giving a final concentration of DMSO of approximately 1% and a final concentration of the complex diluted about 100 times. Attached cells were exposed to the compounds for a 24 h period. Cell respiration, as an indicator of cell viability, was then determined by the mitochondrial-dependent reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) to formazan.\[23\] MTT solution (0.5 mg mL−1) was added to cell cultures and incubated for 3 h, after
which 100 mL of isopropanol was added to dissolve the precipitated formazan crystals. The conversion of MTT to formazan by metabolically viable cells was monitored in an automated microplate reader at 570 nm. The cell viability percentage was calculated by dividing the average absorbance of the cells treated with the test compounds by that of the control; cell viability percentage was plotted against drug concentration (logarithmic scale) to determine the drug concentration at which 50% of the cells are viable relative to the control (IC50), the error being estimated for the average of 3 trials.

Synthetic procedures

All chemicals used were of analytical grade. The reagents, with high purity, were from Sigma-Aldrich Inc., (St. Louis, USA). First, 0.15 mmol of Et3N was added to a degassed solution of 0.10 mmol of cis-[RuCl2(P-P)(bipy)] (P-P = dppm, dppe, dppp or dppf)44 in 30 mL of CH2Cl2. The mixture was allowed to react for 24 h under argon then 0.15 mmol of NH4PF6 was added in order to replace the chloride counter-ion, by the PF6−. After one hour of reaction, the volume of the mixture was reduced to ca. 2 mL and the complexes were precipitated by addition of 15 mL of diethyl ether. The precipitate was filtered off, washed with water (2 × 10 mL) and diethyl ether (2 × 10 mL). Yield: 85-95%.

[Ru(pic)(dppm)(bipy)]PF6(1)

NMR 1H (400 MHz, (CD3)2CO) δ 9.00 (m, 1H, bipy), 8.47 (d, 1H, J 8.4 Hz, bipy), 8.38 (d, 1H, J 5.6 Hz, bipy), 8.35 (d, 1H, J 8.4 Hz, bipy), 8.14 (dt, 1H, J 5.6 and 1.6 Hz, bipy), 8.08-8.04 (m, 2H, Ph), 8.02-7.91 (1H, pic; 2H, phenyl (Ph); m, 1H, bipy), 7.78 (t, 1H, J 6.4 Hz, pic), 7.68-7.51 (1H; pic, m, 6H, Ph), 7.50-7.45 (m, 2H, Ph), 7.29 (t, 1H, J 7.2 Hz, bipy), 7.22 (d, 1H, J 7.2 Hz, bipy), 7.19-6.99 (1H; pic, m, 6H, Ph), 6.81-6.74 (m, 2H, Ph), 5.40-5.28 (m, 1H, CH2), 5.01-4.90 (m, 1H, CH2); anal. calcd. for C33H30N3O3P3F6Ru: exptl./calcd.: C, 53.79/54.09; H, 4.43/4.22; N, 4.66/4.40.

[Ru(pic)(dppe)(bipy)]PF6(2)

NMR 1H (400 MHz, (CD3)2CO) δ 8.95 (m, 1H, bipy), 8.34 (d, 1H, J 8.0 Hz, bipy), 8.17 (d, 1H, J 5.6 Hz, bipy), 8.16-8.08 (4H, m, Ph), 8.02 (dt, 1H, J 7.6 and 1.2 Hz, pic), 7.98-7.94 (m, 1H, bipy), 7.83 (dt, 1H, J 7.6 and 1.2 Hz, pic), 7.72 (t, 1H, J 4.8 Hz, bipy), 7.60-7.50 (1H, bipy; m, 4H, Ph), 7.50-7.41 (m, 5H, Ph), 7.34-7.28 (m, 1H, bipy), 7.26-7.17 (1H; pic, m, 3H, Ph), 7.13 (dt, 1H, J 6.4 and 1.2 Hz, pic), 7.08 (m, 1H, bipy), 6.89 (dt, 2H, J 7.6 and 2.4 Hz, Ph), 6.43 (t, 2H, J 7.2 Hz, Ph), 3.32-2.90 (m, 2H, CH2), 2.50-2.34 (m, 1H, CH2), 2.13-1.98 (m, 1H, CH2); anal. calcd. for C35H33N3O3P3F6Ru: exptl./calcd.: C, 54.48/54.67; H, 3.79/3.93; N, 4.77/4.55.

[Ru(pic)(dppp)(bipy)]PF6(3)

NMR 1H (400 MHz, (CD3)2CO) δ 9.43 (d, 1H, J 5.6 Hz, bipy), 8.66 (m, 1H, bipy), 8.38 (dd, 1H, J 8.4 and 1.2 Hz, bipy), 8.22 (dt, 1H, J 8.4 and 1.2 Hz, bipy), 8.06 (d, 1H, J 8.4 Hz, bipy), 8.02 (m, 2H, Ph), 7.88-7.78 (1H, pic; m, 2H, Ph), 7.75 (dt, 1H, J 7.6 and 1.6 Hz, pic), 7.57-7.42 (m, 6H, Ph), 7.33-7.28 (1H, m, bipy), 7.27-7.17 (1H; pic, m, 1H, bipy), 7.10-7.04 (m, 6H, Ph), 6.98-6.90 (1H, pic; m, 2H, Ph), 6.77 (m, 1H, bipy), 6.38 (t, 2H, J 8.0 Hz, Ph), 3.20-3.02 (m, 2H, CH2), 2.79-2.62 (m, 2H, CH2), 2.40-2.20 (m, 1H, CH2), 2.03-1.87 (m, 1H, CH3); anal. calcd. for C35H33N3O3P3F6Ru: exptl./calcd.: C, 53.79/54.09; H, 4.43/4.22; N, 4.66/4.40.

[Ru(pic)(dppf)(bipy)]PF6(4)

NMR 1H (400 MHz, (CD3)2CO) δ 8.89 (d, 1H, J 5.6 Hz, bipy), 8.05-7.98 (m, 2H, bipy), 7.96 (d, 1H, J 8.0 Hz, bipy), 7.91 (dt, 1H, J 7.6 and 1.2 Hz, pic), 7.70 (dt, 1H, J 8.0 and 1.2 Hz, bipy), 7.67-7.61 (m, 2H, Ph), 7.60-7.50 (1H; pic, m, 3H, Ph), 7.39 (dt, 2H, J 7.6 and 1.6 Hz, Ph), 7.35-7.31 (m, 1H, bipy), 7.28-7.20 (m, 4H, Ph), 7.18-7.11 (1H; pic, m, 2H, Ph), 7.05-6.93 (1H; pic, 1H; bipy, m, 5H, Ph), 6.56 (t, 2H, J 6.8 Hz, Ph), 5.24 (m, 1H, cyclopentadienyl (Cp)), 4.79 (m, 1H, Cp), 4.61 (m, 1H, Cp), 4.45 (m, 2H, Cp), 4.42 (m, 1H, Cp), 4.36 (m, 1H, Cp), 4.16 (m, 1H, Cp); anal. calcd. for C35H33N3O3P3F6Ru: exptl./calcd.: C, 54.10/54.31; H, 3.66/3.73; N, 4.06/3.79.

Results and Discussion

Synthesis and characterization

Complexes 1-4 were obtained from the precursors cis-[RuCl2(P-P)(bipy)] [P-P = 1,1-bis(diphenylphosphino) methane (dppm, 1), 1,2-bis(diphenylphosphino) ethane (dppe, 2), 1,3-bis(diphenylphosphino)propane (dppp, 3) or 1,1′-bis(diphenylphosphino)ferrocene (dppf, 4)] by two chlorido ligands exchanged to one picolinate ion (see Scheme 1). The molar conductivity measurements for the ruthenium complexes (1-4), in dichloromethane, present values consistent with 1:1 electrolytes (17.1-36.2 S cm−1 mol−1), in which one PF6− anion is present as counter-ion. 31P{1H} NMR spectra of the compounds 1-4, 161.98 MHz, CD2Cl2/D2O, show a pair of doublets in a typical AX spin system (Table 1 and Figure S1, Supplementary Information (SI) section), indicating the magnetic non-equivalence of the two phosphorus atoms from the diphosphines.
Table 1. $^{31}P$ [$^1H$] NMR (161.98 MHz, CH$_2$Cl$_2$/D$_2$O), electrochemical data and HOMO’s energy of the compounds

<table>
<thead>
<tr>
<th>Complex</th>
<th>$\delta$ $^{31}P$ [$^1H$] / ppm (2$^1J_{PP}$/ Hz)</th>
<th>$E_{pa}$ / mV</th>
<th>$I_{pa}/I_{pc}$</th>
<th>$E_{HOMO}$ / (kJ mol$^{-1}$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>14.9; 11.9 (60.3)</td>
<td>1277</td>
<td>0.99</td>
<td>$-762.00$</td>
</tr>
<tr>
<td>2</td>
<td>67.6; 65.1 (19.0)</td>
<td>1321</td>
<td>1.01</td>
<td>$-765.04$</td>
</tr>
<tr>
<td>3</td>
<td>35.1; 30.8 (46.9)</td>
<td>1206</td>
<td>0.92</td>
<td>$-761.89$</td>
</tr>
<tr>
<td>4</td>
<td>44.5; 37.8 (32.9)</td>
<td>822$^{a}$/1550</td>
<td>0.89$^{a}$</td>
<td>$-754.33$</td>
</tr>
<tr>
<td>cis-[RuCl$_2$(dppm)(bipy)]</td>
<td>18.5; 11.2 (64.3)</td>
<td>680</td>
<td>1.06</td>
<td>$-442.05$</td>
</tr>
<tr>
<td>cis-[RuCl$_2$(dppe)(bipy)]</td>
<td>68.0; 61.0 (54.0)</td>
<td>726</td>
<td>0.98</td>
<td>$-456.84$</td>
</tr>
<tr>
<td>cis-[RuCl$_2$(dppp)(bipy)]</td>
<td>37.7; 29.8 (42.1)</td>
<td>690</td>
<td>0.92</td>
<td>$-453.19$</td>
</tr>
<tr>
<td>cis-[RuCl$_2$(dppf)(bipy)]</td>
<td>41.8; 36.2</td>
<td>695/1032$^a$</td>
<td>1.350/0.99$^a$</td>
<td>$-456.42$</td>
</tr>
</tbody>
</table>

$^a$Refers to Fe$^{II}$/Fe$^{III}$ couple. $\delta$: chemical shift; $^1J_{PP}$: P-P coupling constant; $E_{pa}$: anodic peak potential; $I_{pa}/I_{pc}$: anodic peak current/cathodic peak current ratio; dppm: bis(diphenylphosphino)methane; dppe: 1,2-bis(diphenylphosphino)ethane; dppf: 1,1’-bis(diphenylphosphino)ferrocene; HOMO: highest occupied molecular orbital.

$^{31}P$ [$^1H$] and $^1H$ NMR studies

All complexes present heptets at $-144$ ppm, corresponding to the PF$_6^-$ counter-ion. The free ligands dppm, dppp, dppf and dppe show $^{31}P$ [$^1H$] chemical shift at $-22.0$, $-17.7$, $-16.5$ and $-11.7$ ppm, respectively. Previously, we assigned the doublet in a higher frequency region to the phosphorus atoms with shorter Ru–P distances. This assignment could be further reinforced in the kinetics studies on the coordination of benzonitrile to cis-[RuCl$_2$(dppb)(phen)] followed by the $^{31}P$ [$^1H$] NMR experiment reported elsewhere.

Interestingly, the chemical shift of the complex containing the dppe ligand is very high in comparison to the values observed for the other compounds. This can be explained in terms of the “ring contribution” which is mentioned as the factor responsible for the high values of chemical shifts observed and are caused by the unshielded associated to the formation of the very stable five-membered ring. The stability of the complexes in the biological medium was checked by $^{31}P$ [$^1H$] NMR experiments, and the complexes showed to be stable in this solution for at least three days.

Concerning the $^1H$ NMR spectra of complexes 1-4, signals at 8.16-6.38 ppm correspond to twenty hydrogen atoms of the phenyl groups of the diphosphine ligands occur as a series of multiplets, while the eight protons of the bipy ligand can be observed in the typical region, in the range of 9.43-6.77 ppm. The coordination of the picolinate ligand is verified by the presence of four signals, corresponding to the four hydrogen atoms, in the region of 8.02-6.90 ppm. In complex 4, due to the non-equivalence of the phosphorus atoms and the conformation of the cyclopentadienyl (Cp) rings, an asymmetric distribution of electronic density.

Scheme 1. Route for synthesis of the complexes 1-4.
occurs and the spectrum of this compound shows signals in the 5.24–4.16 ppm region corresponding to the eight protons of Cp rings. The protons of the −CH− groups of the diphosphines are observed as multiplets in 3.20–1.87 ppm (six hydrogen atoms), 3.32–1.98 ppm (four hydrogen atoms) and 5.40–4.90 ppm (two hydrogen atoms) regions for compounds 3, 2 and 1, respectively (see Figure S2, SI section).

Structural studies

As expected, all four complexes presented distorted octahedral coordination geometry around the ruthenium center. The crystal structures of complexes 1–4 were determined (Figure 1) and the selected bond distances and angles are presented in Table 2. Data collection and experimental details are summarized in Table S1 (see SI section). The coordination of 2-picolinate to the ruthenium center formed a five-member ring with N1−Ru−O1 angle values close to those ones found N2−Ru−N3 bipyridine chelate ring, presenting angle values around 78º for all the complexes. The bite angle measured based on P1−Ru−P2 bond angles are 71.63(14), 82.64(3), 92.43(3) and 97.19(3) for the complexes containing the diphosphines dpdm (1), dppe (2) and dppp (3) and dppf (4), respectively. Calculations showed N2−Ru−N3 around 77º, for all four complexes and P1−Ru−P2 equal to 72.82, 83.90, 91.24 and 97.50º for 1, 2, 3 and 4, respectively.

The structures of 2 and 4 have one molecule of water, each, in their asymmetric unities, which make hydrogen bonds with the fluorine atoms from the PF6− anion and with the oxygen from picolinate ligand. The distances H⋯F are equal to 2.541 and 2.620 Å, and H⋯O equal to 2.037 and 1.909 Å in the complexes 2 and 4, respectively. The non-existence of individual bonds between the metal and the Cp rings, gives some mobility to the rings, allowing them to rotate with respect to each other.40 As a result of the coordination of the diphosphine to the ruthenium metal in a bidentate way, as well as the relatively high “bite angle” of dppf and the difference in the Ru−P bond lengths, the Cp rings are not parallel to each other. In fact, they form an angle of 2.36º and present torsion angles P−Cp−Cp−P of 22.99º in complex 4, which is very similar to the cis-[RuCl2(dppe)(bipy)] that presents a torsion angle of 19.70º,34 including them in a “synclinal staggered” conformation, such as previously discussed by Bandoli and Dolmella.41

As expected, the C−O (coordinated to ruthenium) bond lengths (ca. 1.30 Å) are longer than the C=O distances. In the molecule of the free picolinic acid, the C=O and C−OH distances are equal to 1.214 and 1.278 Å, respectively.42 The Ru−N1 distances do not change considerably with the exchange of the diphosphines, as can be seen in Table 2. The distances of Ru−P and Ru−Nbipy bonds in the new complexes (Table 2) are shorter than in precursor complexes, indicating a more efficient back-donation from ruthenium to phosphorus atoms in the precursor complexes. In the cis-[RuCl2(dppe)(bipy)] complex, the Ru−P distance is equal to 2.2465(8) Å for the phosphorus atoms trans positioned to the chlorido ligand and 2.2907(9) Å for the phosphorus atoms trans positioned to the nitrogen atom from the bipy ligand,35 while in 2 these distances are equal to 2.2998(9) Å for Ru−P1 and 2.3166(9) Å for Ru−P2.

Table 2. Selected bond distances and bond angles of complexes 1–4

<table>
<thead>
<tr>
<th>Fragment</th>
<th>Bond distance / Å</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ru−P1</td>
<td>2.285(4)</td>
<td>2.2998(9)</td>
<td>2.3149(8)</td>
<td>2.3480(7)</td>
<td></td>
</tr>
<tr>
<td>Ru−P2</td>
<td>2.302(4)</td>
<td>2.3166(9)</td>
<td>2.3270(8)</td>
<td>2.3515(7)</td>
<td></td>
</tr>
<tr>
<td>Ru−N1</td>
<td>2.129(11)</td>
<td>2.131(3)</td>
<td>2.132(2)</td>
<td>2.118(2)</td>
<td></td>
</tr>
<tr>
<td>Ru−N2</td>
<td>2.108(12)</td>
<td>2.111(3)</td>
<td>2.105(2)</td>
<td>2.106(2)</td>
<td></td>
</tr>
<tr>
<td>Ru−N3</td>
<td>2.056(12)</td>
<td>2.073(3)</td>
<td>2.076(2)</td>
<td>2.062(2)</td>
<td></td>
</tr>
<tr>
<td>Ru−O1</td>
<td>2.094(9)</td>
<td>2.087(2)</td>
<td>2.106(2)</td>
<td>2.094(2)</td>
<td></td>
</tr>
<tr>
<td>C1−O1</td>
<td>1.272(17)</td>
<td>1.291(4)</td>
<td>1.275(4)</td>
<td>1.260(4)</td>
<td></td>
</tr>
<tr>
<td>C1=O2</td>
<td>1.219(18)</td>
<td>1.226(5)</td>
<td>1.223(4)</td>
<td>1.233(4)</td>
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<table>
<thead>
<tr>
<th>Bond angle / degree</th>
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<tr>
<td>P1−Ru−P2</td>
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<tr>
<td>N2−Ru−N3</td>
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<tr>
<td>N1−Ru−O1</td>
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<tr>
<td>C1−O1</td>
</tr>
<tr>
<td>C1=O2</td>
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</table>

Figure 1. Crystal structure of the complexes 1, 2, 3 and 4 showing selected atoms labeling and the ellipsoids at 30% probability. PF6− counter-ion was omitted for clarity.
As it can be seen in Table 2, Ru–P back donation is more efficient in the precursors complexes, because the strong electron σ- and π-donor character of the chlorido ligand enrich the ruthenium center of electron density, facilitating the Ru–P backdonation. The distances Ru–P and Ru–N(bipy) are in the range usually reported in the literature for complexes containing diphosphines. 36,37 The packing effect must be considered because experimental measurements are made in solid state and also counter ion and solvent presence help contracting distances. These factors alter intermolecular dispersive and electrostatic forces creating a potential energy around molecules influencing their geometry. 38

Electrochemical, infrared and UV-Vis studies for 1-4

Cyclic voltammograms of compounds 1, 2 and 3 show one reversible process, with \( E_{pa} \) (anodic peak potential) equal to 1277, 1321 and 1206 mV, respectively (Table 1 and Figures S3-S6, SI section), while the precursor complexes present \( E_{pa} \) values, around 680, 726 and 690 mV, for the compounds cis-[RuCl\(_2\)(dppm)(bipy)] \( \), cis-[RuCl\(_2\)(dppe)(bipy)] \( \) and cis-[RuCl\(_2\)(dpff)(bipy)] \( \), respectively. Complex 4 presents one quasi-reversible process with \( E_{pa} \) equal to 822 mV, attributed to the Fe\(^{II}/Fe^{III} \) redox pair and one irreversible process with \( E_{pa} \) equal to 1550 mV corresponding to the Ru\(^{II}/Ru^{III} \) pair. Similarly, its precursor cis-[RuCl\(_2\)(dppf)(bipy)] \( \) presents two reversible redox pairs with \( E_{pa} \) values equals to 685 and 1032 mV, but in this case the former corresponds to Fe\(^{II}/Fe^{III} \) and Ru\(^{II}/Ru^{III} \), respectively. 44,45 Electrochemical attribution of the redox pairs to the Ru\(^{II}/Ru^{III} \) or Fe\(^{II}/Fe^{III} \) process in the voltammogram of 4 was made as follows: exhaustive electrolysis at an intermediate potential (900 mV) was performed and then a few drops of a NH\(_4\)SCN solution were added to the electrochemical cell, and immediately a characteristic red color appears indicating the formation of [Fe(SCN)\(_6\)]\(^{3-} \), which confirms the presence of Fe\(^{III} \). Due to the coordination of the picolinate ligand to the metal center, the \( E_{pa} \) values increase making complexes 1-4 more stable than cis-[RuCl\(_2\)(P-P)(bipy)]. This behavior can be explained because picolinate is a better σ-donor and π-acceptor ligand than the chlorido ligands, which acts as σ- and π-donor. When the π-acceptor character of the ligand decreases, it is expected that the value of \( E_{pa} \) also decreases, such as observed for the complexes [Ru(bipy)]\(_2\)\(^{3+} \) (\( E_{pa} = 1300 \) mV), 46 [Ru(pic)(bipy)]\(_3\)\(^{+} \) (\( E_{pa} = 750 \) mV), 47 [Ru(pic)\(_3\)]\(^{2+} \) (\( E_{pa} = 440 \) mV) and [Ru(pic)\(_2\)] (\( E_{pa} = -90 \) mV). 48 From these data, it can be concluded that the π-acceptor character of the picolinate ligand is weaker than that of the bipy ligand.

In addition, the diphosphines are better π-acceptor than bipy, according to the oxidation potentials observed for the complexes synthesized herein and [Ru(pic)(bipy)]\(_2\)ClO\(_4\) (\( E_{pa} = 750 \) mV). The substitution of one bipy in [Ru(pic)(bipy)]\(^{+}\) by diphosphines yielded the complexes studied here and resulted in higher \( E_{pa} \) values of 1277, 1321, 1206 and 1550 mV for 1, 2, 3 and 4, respectively (see Table 1). 46 Theoretical results support these experimental data, in which the replacement of the two chlorido atoms by the picolinate anion stabilizes the complexes. As it is well known, oxidation potential can be related to ionization potential (\( E' \alpha E_{HOMO} \)) since both refer to the withdraw of electrons, which in turn can be associated to the highest occupied molecular orbital (HOMO) energy (\( E_{HOMO} \)). Therefore, oxidation potential can be related to the HOMO energy, i.e., the lower the HOMO energy, the higher the oxidation potential.

As can be seen in Table 1, replacement of the chloride atoms from the precursor complexes (\( E_{HOMO} \) around \(-450 \) kJ mol\(^{-1}\)) decreases the HOMO energies in 1, 2, 3 and 4 (\( E_{HOMO} \) around \(-760 \) kJ mol\(^{-1}\)), consequently the oxidation potentials of the complexes increase.

Charge decomposition analysis (CDA) revealed that charge transference from chlorido ligands to the ruthenium atom is greater in the precursor complexes than in 1-4 (Table S2, SI section). For the cis-[RuCl\(_2\)(dpff)(bipy)] complex, for instance, both chlorido atoms together donate 1.427 e- to the metal center, but the picolinate in the complex 1 donates only 0.695 e-, which makes the ruthenium center charge more negative in the precursor complexes and explains its lower oxidation potential observed. HOMO energies are located mainly on ruthenium atoms (60%) and chlorido ligands (35%) in cis-[RuCl\(_2\)(P-P)(bipy)], while after the coordination of picolinate anion to the metal center the HOMO is mostly concentrated on ruthenium atoms and on the oxygen atoms. In complexes 1, 2 and 3, contributions of “d” orbitals of the ruthenium to HOMO is around 70% and “p” orbitals of oxygen atoms are around 20%. In 4, HOMO has 42% contribution of “d” orbitals of the ruthenium and 35% of “d” orbitals of iron (Figure 2) in its precursor. On the other hand, 59% of HOMO comprises ruthenium “d” orbitals and iron orbitals have a negligible participation in it. That was expected, since the first oxidation potential refers to the Ru\(^{II}/Ru^{III} \) process in cis-[RuCl\(_2\)(dpff)(bipy)] and to Fe\(^{II}/Fe^{III} \) in 4.

In the infrared spectra the calculated and experimental vibrational data are compared. All complexes presented the expected bands relative to \( \nu_\alpha \)C–H and \( \nu_\gamma \)C–H of –CH\(_2\), picolinate, bipy and phenyl moieties at around 3060-2900 cm\(^{-1}\) region. For complex 4, bands attributed
to $\nu_{\text{as}}\text{C}^{-}\text{H}$ and $\nu_{s}\text{C}^{-}\text{H}$ of Cp rings were observed in the same region, instead of $-\text{CH}_2$. The bands at around 1610-1400 cm$^{-1}$ region were attributed to C=C stretching. Additional bands were observed in 1400-990 cm$^{-1}$ and 990-600 cm$^{-1}$ regions, the former corresponding to C-H stretching in the plane and the latter out of the plane. Bands at 840 and 557 cm$^{-1}$ revealed the presence of the counter-ion PF$_6^{-}$. In complexes 1-4, the bands corresponding to $\nu_{\text{as}}(\text{COO})$ and $\nu_{s}(\text{COO})$ appeared at ca. 1655 and 1335 cm$^{-1}$, respectively, whereas in the free ligand these bands can be found at 1654 and 1350 cm$^{-1}$, respectively. The difference between $\nu_{\text{as}}(\text{COO})$ and $\nu_{s}(\text{COO})$ coordinated to the metal centers is an indicative of the type of coordination. In this case $\nu_{\text{as}}(\text{COO})-\nu_{s}(\text{COO})$ ca. 320 cm$^{-1}$ it shows that the carboxylate group is coordinated to ruthenium in a monodentate way. Also, bands at 1600 cm$^{-1}$ related to $\nu$(C=N) were observed. For complex 4, weak bands corresponding to $\nu$(Fe-Cp) and angular deformation of C-H bond of the cyclopentadienyl ring were observed at 546 and 1041 cm$^{-1}$, respectively. For the complexes containing the diphosphines dppe and dppe weak bands corresponding to $\nu$(P=CH$_2$) at 667, 676 and 669 cm$^{-1}$ were observed, respectively. The infrared (IR) spectra and their assignments for complexes 1-4 are shown in the SI section (Table S3 and Figure S7).

The UV-Vis spectra of compounds 1-4 are quite similar to each other (Figure S8 and Table S4, SI section). All complexes presented bands at 300 and 420 nm and shoulders around 340 and 500 nm. The bands in the UV region, centered at 300 nm are attributed to $\pi\rightarrow\pi^*$ intraligand charge transfer type, which is also observed in the spectra of the free ligands P-P and picH. Calculations support the attributions of the transitions. The bands in the region 340 (shoulder), 420 and 500 (shoulder) nm can be assigned to a metal to ligand charge transfer (MLCT).

Cytotoxicity of 1-4 against MDA-MB-231 and murine ascistic sarcoma 180 tumor cells

Evaluation of the cytotoxicity for the complexes 1-4 against MDA-MB-231 (human breast carcinoma) and S180 (murine ascistic sarcoma 180) tumor cells, after incubation of 24 h, were carried out. The IC$_{50}$ values were calculated using the dose-survival curves with MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay. Complexes 1-4 present very good activity against the MDA-MB-231 cell line (Table 3), with values of IC$_{50}$ lower than cisplatin indicating their potentiality as antitumor agents. The complexes were also active against murine sarcoma, presenting IC$_{50}$ values lower than 72 μM, while for cisplatin the IC$_{50}$ value against the S180 cell line is 64.83 ± 0.17 μM. For complexes 1, 2, 3 and 4, the IC$_{50}$ values against S180 are equal to 71.9, 31.3, 11.2 and 3.5 μM, respectively. Meanwhile the P–Ru–P bite angles are 71.63(14), 82.64(3), 92.43(3), 97.19(3) Å. It is worth mentioning that the IC$_{50}$ values decrease concomitantly with the diphosphine bite angle increase. This aspect suggests a relationship between these parameters. The same tendency is also observed against MDA-MB-231 tumor cells (see Table 3).

<table>
<thead>
<tr>
<th>Complex</th>
<th>IC$_{50}$ values / μM</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>S180</td>
</tr>
<tr>
<td>1</td>
<td>71.91 ± 0.13</td>
</tr>
<tr>
<td>2</td>
<td>31.32 ± 0.19</td>
</tr>
<tr>
<td>3</td>
<td>11.21 ± 0.15</td>
</tr>
<tr>
<td>4</td>
<td>3.53 ± 0.21</td>
</tr>
<tr>
<td>Cisplatin</td>
<td>64.83 ± 0.17</td>
</tr>
</tbody>
</table>

IC$_{50}$: drug concentration at which 50% of the cells are viable relative to the control.

Lipophilicity or partition coefficient is usually expressed by log P, which describes the equilibrium between water and an immiscible lipid-like organic solvent, for example n-octanol. Log P is the ratio of concentrations in the two phases $[\log P = [\text{metalodrug}]_{\text{octanol}} / [\text{metalodrug}]_{\text{water}}]$, so that a positive value for log P reflects a preference for the lipid phase, and a negative value reflects the relative affinity of the complex for water. Complexes 1-4 showed
log P positive, suggesting a preference for the lipid phase and possibly allowing permeation across biological membranes. The calculated log P showed positive values for complexes 1 (0.95 ± 0.15) and 2 (1.22 ± 0.11), complex 3 (1.31 ± 0.12), and complex 4 (1.54 ± 0.11). The order of hydrophobicity in nature of the complexes is 1 < 2 < 3 < 4. Hydrophobicity is one of the most important properties to express the biological activity of the compounds, since it is necessary to cross biological membranes in order to reach their respective sites of action. Hydrophobic compounds can increase cell uptake and improve their anticancer activity.

The new compounds showed better activity than the precursor ones, except for compound 1, suggesting that the presence of picolinate ligand might play an important role in the activity of the complexes. Interestingly, the free ligands, including the picolinic acid, are not active against the tumor cells tested herein, in which the IC_{50} values are higher than 200 μM. It is important to point out that the precursor complexes [RuCl_{2}(P-P)(bipy)] may act as pro-drugs, given that after their dissolution in DMSO, to prepare stock solution, there is a rapid exchange of one chloride ligand, reacting with DMSO. A number of complexes reported in the literature have their antitumor activity associated to a dissociation of chloride ligand, suggesting that the mechanism possibly involves covalent interactions with DNA, which probably does not occur with the stable hexacoordinated compounds, presenting three bidentate ligands (bipy, pic, P-P) reported herein. It is worth mentioning that the complexes 1-4 are stable in DMSO solutions for at least 5 days, which was showed by ^{31}P NMR experiments. Instead, a possible mechanism to explain the activity might be a non-covalent interaction of ruthenium compounds with the DNA. Moreover, it is important to point out that DNA is not the only target aimed for antitumor agents. Further investigations are necessary to help the elucidation of the mechanism of action of the complexes.

**Conclusions**

In this work, four new complexes [Ru(pic)(P-P)(bipy)]PF_{6}, pic = picolinate, P-P = dppf, dppp, dppe and dppm were synthesized, characterized by spectroscopic, electrochemical and X-ray crystallography techniques and had their structures elucidated theoretically. X-ray data showed that the phosphorus atoms from the diphosphine are trans positioned to the nitrogen atoms from the bipy and pic ligands. The compounds had their cytotoxicity evaluated in vitro against the human tumor cell lines MDA-MB-231 and murine ascitic sarcoma 180 showing good activities. Among the new compounds, the [Ru(pic)(dppf)(bipy)]PF_{6} presented the highest activity against both cells tested, followed by [Ru(pic)(dppp)(bipy)]PF_{6}, [Ru(pic)(dppe)(bipy)]PF_{6} and [Ru(pic)(dppm)(bipy)]PF_{6}, which may indicate that the complexes have the same mechanism of action against both MDA-MB-231 and S180 human tumor cell lines. Furthermore, higher activities were observed for those complexes containing diphosphines dppf and dppp than those containing dppe and dppm suggesting a relationship between bite angle and activity. The lipophilicity of the complexes influenced the antiproliferative activity of the cells. The most lipophilic complex was the most active. However, further investigations are necessary in order to help the elucidation of the mechanism of action of the complexes.

**Supplementary Information**

Crystallographic data (excluding structure factors) for the structures in this work were deposited in the Cambridge Crystallographic Data Centre as supplementary publication numbers for complex 1 (1912057), complex 2 (1815880), complex 3 (1815881) and complex 4 (1912056). Copies of the data can be obtained, free of charge, via www.ccdc.cam.ac.uk/conts/retrieving.html or from the Cambridge Crystallographic Data Centre, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033. E-mail: deposit@ccdc.cam.ac.uk.

Supplementary information (the spectra and tables for 1-4) is available free of charge at http://jbcs.sbq.org.br as PDF file.

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**Author Contributions**

Augusto Lima was responsible for the software; Rodrigo Correa for the investigation, writing-review and editing; Angelica Graminha for the investigation; Jaldyr Jesus Varela for the investigation and software; Alberico da Silva for the investigation and software; Javier Ellena for the investigation; Thales Eduardo Silva for the investigation; Alzir A. Batista for the conceptualization and supervision.
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