Novel Non Enzymatic TBHQ Modified Electrochemical Sensor for Hydrogen Peroxide Determination in Different Beverage Samples

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Table S1. Comparison of the electrocatalytic reduction peak potential (E_p) and peak current (I_p) of $H_2O_2(0.3 \text{ mmol } L^{-1})$ on various electrode surfaces at pH 7.0

Name of electrode	Reduction potential / mV	Reduction current / µA
N-GO/TBHQ/GCE ^a	-142	-0.10
AgNPs/TBHQ/GCE ^b	-109	-0.12
N-GO/AgNPs/TBHQ/GCE ^c	-97	-0.70

^aN-GO/TBHQ/GCE: nafion, graphene oxide and TBHQ modified glassy carbon electrode; ^bAgNPs/TBHQ/GCE: silver nanoparticles and TBHQ modified glassy carbon electrode; ^cN-GO/AgNPs/TBHQ/GCE: nafion, graphene oxide, silver nanoparticle and TBHQ modified glassy carbon electrode.



Figure S1. Cyclic voltammetric responses of N-GO/AgNPs/TBHQ/GCE in 0.1 mol L⁻¹ phosphate buffer (pH 7.0) at different scan rates (5-95 mV s⁻¹). Insets: (a) plots of anodic and cathodic peak currents *vs.* scan rate; (b) variation of the peak potentials *vs.* the logarithm of the scan rate; (c) magnification of the same plot for high scan rates.

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Figure S2. (a) Plot of the electrocatalytic currents vs. the square root of scan rate; (b) linear sweep voltammogram recorded at 14-24 mV s⁻¹. The conditions are the same as in (a). Inset shows the Tafel plot derived from the linear sweep voltammogram.



Figure S3. Chronoamperometric response at an N-GO/AgNPs/TBHQ/GCE in 0.1 mol L⁻¹ phosphate buffer (pH 7.0) at a potential step of -150 mV for different concentrations of hydrogen peroxide. The numbers 1-7 correspond to 4, 5, 10, 20, 30, 40, and 50 mmol L⁻¹ of H₂O₂. Insets: (a) plots of I vs. t^{-1/2} obtained from the chronoamperograms; (b) plot of the slope of the straight lines against the hydrogen peroxide concentration.



Figure S4. 8 reparative cyclic voltammograms of the N-GO/GCE with continuous potential cycling from -0.7 to 1.9 V at a sweep rate of 80 mV s⁻¹ in a solution containing 1 mmol L⁻¹ AgNO₃ and 100 mmol L⁻¹ nitric acid.